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(54) Title: HETEROLOGOUS G-CSF FUSION PROTEINS

(57) Abstract: The present invention encompasses heterologous fusion proteins comprising a hyperglycosylated G-CSF analog fused to proteins such as albumin and the Fc portion of an immunoglobulin which act to extend the in vivo half-life of the protein compared to native G-CSF. These fusion proteins are particularly suited for the treatment of conditions treatable by stimulation of circulating neutrophils, such as after chemotherapy regimens or in chronic congenital neutropenia.

HETEROLOGOUS G-CSF FUSION PROTEINS

The present invention relates to heterologous fusion proteins, including analogs and derivatives thereof, fused to proteins that have the effect of extending the *in vivo* half-life of the proteins. These fusion proteins are significant in human medicine, particularly in the treatment of conditions treatable by stimulation of circulating neutrophils, such as after chemotherapy regimens or in chronic congenital neutropenia. More specifically, the invention relates to novel heterologous fusion proteins with granulocyte-colony stimulating factor activity.

Among all blood cell lineages, the modulation of neutrophil and platelet production has been of highest interest to clinical oncologists and hematologists. Myelosuppression is the single most severe complication of cancer chemotherapy, and a major cause of treatment delay during multiple-cycle or combination chemotherapy. It is also the major dose-limiting factor for most chemotherapeutic agents. Due to the short half-lives of neutrophils in peripheral blood, life-threatening falls in neutrophil levels are seen after a number of conventional anti-tumor chemotherapy regimens.

The most prominent regulator of granulopoiesis is granulocyte-colony stimulating factor (G-CSF). G-CSF induces proliferation and differentiation of hematopoietic progenitor cells resulting in increased numbers of circulating neutrophils. G-CSF also stimulates the release of mature neutrophils from bone marrow and activates their functional state. [Souza L.M., et al. (1986) *Science* 232:61-65]. Thus, therapeutic proteins with G-CSF activity have tremendous value in situations where there are reduced circulating levels of neutrophilic granulocytes.

However, the usefulness of therapy using G-CSF peptides has been limited by their short plasma half-life. Thus,

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they must be administered intravenously or subcutaneously at fairly frequent intervals (once or twice a day) in order to maintain their neutrophil stimulating properties. In addition, this short half-life limits the performance of the drug to traditional drug delivery systems. It would clearly benefit the treatment of patients with abnormally low neutrophils, and reduce the discomfort and inconvenience associated with frequent injections to provide a pharmaceutical agent that could be administered less frequently and optionally by alternative routes of administration. Thus, a need exists to develop agents that stimulate the production of mature neutrophils and are more optimal in their duration of effect.

The present invention overcomes the problems associated with delivering a compound that has a short plasma half-life in two respects. First, G-CSF is hyperglycosylated. The carbohydrate content of G-CSF is altered by substituting amino acids that can act as substrates for glycosylating enzymes in mammalian cells. Most significantly, the present invention encompasses fusion of these hyperglycosylated G-CSF analogs to another protein with a long circulating half-life such as the Fc portion of an immunoglobulin or albumin.

Compounds of the present invention include heterologous fusion proteins comprising a hyperglycosylated G-CSF analog fused to a polypeptide selected from the group consisting of
a) human albumin;
b) human albumin analogs; and
c) fragments of human albumin.

Compounds of the present invention also include heterologous fusion proteins comprising a hyperglycosylated G-CSF analog fused to a polypeptide selected from the group consisting of
a) human albumin;
b) human albumin analogs; and
c) fragments of human albumin,

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wherein the hyperglycosylated G-CSF analog is fused to the polypeptide via a peptide linker.

Additional compounds of the present invention include a heterologous fusion protein comprising a hyperglycosylated G-CSF analog fused to a polypeptide selected from the group consisting of

- a) the Fc portion of an immunoglobulin;
- b) an analog of the Fc portion of an immunoglobulin;
and
- c) fragments of the Fc portion of an immunoglobulin.

The G-CSF analog may be fused to the polypeptide via a peptide linker. It is preferable that the peptide linker is selected from the group consisting of:

- a) a glycine rich peptide;
- b) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]_n where n is 1, 2, 3, 4, or 5; and
- c) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]₃.

The present invention further provides data showing that these G-CSF analogs are glycosylated in mammalian cells and retain their activity.

One aspect of the present invention includes heterologous fusion proteins, wherein the hyperglycosylated G-CSF analogs have the Formula (I) [SEQ ID NO:1]

1	5	10	15
Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Leu Lys			
20	25	30	
Xaa Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln			
35	40	45	
Glu Lys Leu Cys Xaa Xaa Xaa Lys Leu Cys His Pro Glu Glu Leu Val			
50	55	60	
Leu Leu Gly His Ser Leu Gly Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa			
65	70	75	80
Kaa Xaa Xaa Xaa Xaa Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser			
85	90	95	
Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Xaa Xaa Xaa Ser			
100	105	110	
Kaa Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp			
115	120	125	
Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro			
130	135	140	
Ala Leu Gln Pro Xaa Xaa Xaa Ala Met Pro Ala Phe Xaa Xaa Xaa Phe			
145	150	155	160
Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe			

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165 170
Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro (I)

wherein:

Xaa at position 17 is Cys, Ala, Leu, Ser, or Glu;
Xaa at position 37 is Ala or Asn;
Xaa at position 38 is Thr, or any other amino acid except Pro;
Xaa at position 39 is Tyr, Thr, or Ser;
Xaa at position 57 is Pro or Val;
Xaa at position 58 is Trp or Asn;
Xaa at position 59 is Ala or any other amino acid except Pro;
Xaa at position 60 is Pro, Thr, Asn, or Ser,
Xaa at position 61 is Leu, or any other amino acid except Pro;
Xaa at position 62 is Ser or Thr;
Xaa at position 63 is Ser or Asn;
Xaa at position 64 is Cys or any other amino acid except Pro;
Xaa at position 65 is Pro, Ser, or Thr;
Xaa at position 66 is Ser or Thr;
Xaa at position 67 is Gln or Asn;
Xaa at position 68 is Ala or any other amino acid except Pro;
Xaa at position 69 is Leu, Thr, or Ser
Xaa at position 93 is Glu or Asn
Xaa at position 94 is Gly or any other amino acid except Pro;
Xaa at position 95 is Ile, Asn, Ser, or Thr;
Xaa at position 97 is Pro, Ser, Thr, or Asn;
Xaa at position 133 is Thr or Asn;
Xaa at position 134 is Gln or any other amino acid except Pro;
Xaa at position 135 is Gly, Ser, or Thr
Xaa at position 141 is Ala or Asn;
Xaa at position 142 is Ser or any other amino acid except Pro; and
Xaa at position 143 is Ala, Ser, or Thr;

and wherein:

Xaa at positions 37, 38, and 39 constitute region 1;
Xaa at positions 58, 59, and 60 constitute region 2;
Xaa at positions 59, 60, and 61 constitute region 3;
Xaa at positions 60, 61, and 62 constitute region 4;
Xaa at positions 61, 62, and 63 constitute region 5;

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Xaa at positions 62, 63, and 64 constitute region 6;
Xaa at positions 63, 64, and 65 constitute region 7;
Xaa at positions 64, 65, and 66 constitute region 8;
Xaa at positions 67, 68, and 69 constitute region 9;
Xaa at positions 93, 94, and 95 constitute region 10;
Xaa at positions 94, 95, and Ser at position 96
constitute region 11;
Xaa at positions 95, and 97, and Ser at position 96
constitute region 12;
Xaa at positions 133, 134, and 135 constitute
region 13;
Xaa at positions 141, 142, and 143 constitute
region 14;

and provided that at least one of regions 1 through 14
comprises the sequence Asn Xaa1 Xaa2 wherein Xaa1 is any
amino acid except Pro and Xaa2 is Ser or Thr.

Thus, the heterologous fusion proteins of the present
invention include analogs wherein one or any combination of
two or more regions comprise the sequence Asn Xaa1 Xaa2
wherein Xaa1 is any amino acid except Pro and Xaa2 is Ser or
Thr.

Preferred hyperglycosylated G-CSF analogs that make up part
of the heterologous fusion proteins of the present
invention, include the following:

- a) G-CSF[A37N, Y39T]
- b) G-CSF[P57V, W58N, P60T]
- c) G-CSF[P60N, S62T]
- d) G-CSF[S63N, P65T]
- e) G-CSF[Q67N, L69T]
- f) G-CSF[E93N, I95T]
- g) G-CSF[T133N, G135T]
- h) G-CSF[A141N, A143T]
- i) G-CSF[A37N, Y39T, P57V, W58N, P60T]
- j) G-CSF[A37N, Y39T, P60N, S62T]
- k) G-CSF[A37N, Y39T, S63N, P65T]
- l) G-CSF[A37N, Y39T, Q67N, L69T]
- m) G-CSF[A37N, Y39T, E93N, I95T]
- n) G-CSF[A37N, Y39T, T133N, G135T]
- o) G-CSF[A37N, Y39T, A141N, A143T]

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- p) G-CSF [A37N,Y39T,P57V,W58N,P60T,S63N,P65T]
- q) G-CSF [A37N,Y39T,P57V,W58N,P60T,Q67N,L69T]
- r) G-CSF [A37N,Y39T,S63N,P65T,E93N,I95T]

The present invention also includes heterologous fusion proteins, which are the product of the expression in a host cell of an exogenous DNA sequence, which comprises a DNA sequence encoding a heterologous fusion protein of Formula I (described above) fused to a DNA sequence encoding human albumin or the Fc portion of an immunoglobulin.

The present invention includes an isolated nucleic acid sequence, comprising a polynucleotide encoding a heterologous fusion protein described above. Exemplary isolated nucleic acids of the present invention include isolated nucleic acid sequence comprising a hyperglycosylated G-CSF analog selected from the group consisting of:

- a) SEQ ID NO:2
ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC
- GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CTT CGC GAG GTC
- GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC AGC GTG GGG CTC CTC GAC CAC
- CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TCG AGC
- CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CCC AGC AAC TCG GTT GAG GTA TCG
- GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG
- CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA CGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG
- TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GGC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT GAC CCT TAC CGG GGA
- GCC CTG CAG CCC AAC CAG ACC GGC ATG CGG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TTG GTC TGG CGG TAC GGC CGG AAG CGG AGA CGA AAG
- CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGC GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

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CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC GGG

b) SEQ ID NO:3

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGG GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC ACG GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TCG ACG

CCC AGC CAG GCC CTG CAG CTG GCA GGG TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CCG ACG AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC AAC TCT ACC TTC
CGG GAC GTC CGG TGG GTC CCA CGG TAC GGC CGG AAG TTG AGA TGG AAG

CGC CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GGC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAC

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC GGG

c) SEQ ID NO:4

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCC GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGG GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC ACG GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCC TAG GGG ACC CGA GGG GAC TCG TCG ACG

CCC AGC CAG GCC CTG CAG CTG GCA GGG TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CCG ACG AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC AC TGT GAC GTC GAC CTG CAG CGG CTG

- 8 -

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA
GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG
CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG
CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC GGG

d) SEQ ID NO:5

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC CGG GTC TCG AAG GAC GAG TTC
GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC
GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC AGC GTG GGG CTC CTC GAC CAC
CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT AAC ACT AGC AGC TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA TTG GAC TCC TCG ACG
CCC AGC CAG GGC CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CGG ACC AAC TCG GTT GAG GTA TCG
GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCC GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG
CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA CGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA
GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG
CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG
CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC GGG

e) SEQ ID NO:6

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC CGG GTC TCG AAG GAC GAG TTC
GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CGG CTA CGG CGT CGC GAG GTC
GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC AGC GTG GGG CTC CTC GAC CAC
CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AAT TCC

- 9 -

GAC GAG CCT GTG ACA GAC CCG TAG GGG ACC CGA GGG GAC TCG TTA ACG

ACC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
TGG TCG GTC CGG GAC GTC CAG CCG ACG AAC TCG GTT GAG GTA TCGGGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGGCCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTGTTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGAGCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CCG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAGCAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAGCTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CGC
GAC CTC CAC AGC ATG CGC CAG AAT TCC GTG GAA CGG GTC CGG

f) SEQ ID NO:7

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTCGCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTCGAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC AGC GTG GGG CTC CTC GAC CACCTG CTC GGA CAC TCT CTG GGC ATC GTT AAC GCT ACC CTG AGC AGC TGC
GAC GAG CCT GTC ACA GAC CCC TAG CAA TTG CGA TGG GAC TCG TCG ACGCCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CGG AGC AAC TCG GTT GAG GTA TCGGGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGGCCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTGTTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGAGCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CCG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAGCAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAGCTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGC CAG AAT TCC GTG GAA CGG GTC CGG

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g) SEQ ID NO:8

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TCG ACG

CCC AGC AAC GGC ACC CAG CTG GCA GGC TGC TTG AGC CRA CTC CAT AGC
GGG TCG TTG CGG TGG GTC GAC CGT CCC AGC AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCC GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA CGG TGG AAC CTG TGT GAC GTC GAC CTG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CGG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GCC CGG AAG CGG AGA CGA AAG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCG CGT CCT CCC CAG GAC CAA CGG AGG GTC GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGG CAG AAT TCC GTG GAA CGG GTC CGG

h) SEQ ID NO:9

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TCG ACG

CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC CGT CCC AGC AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG AAC GGG ACC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC TTG CCC TGG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA CGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT GAC CCT TAC CGG GGA

- 11 -

GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CGC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC CGG

i) SEQ ID NO:10

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC ACC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCG TAG GGG ACC CGA GGG GAC TCG TCG AGC

CCC AGC CAG GCC CTG CAG CTC GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC CTC GAC CGT CCG ACC AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CGG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC AAC CAG ACC GCC ATG CCG GCC TTC GCC TCT GCT GTC
CGG GAC GTC GGG TTG GTC TGG CGG TAC GGC CGG AAG CGG AGA CGA AAG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGF CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CGC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC CGG

j) SEQ ID NO:11

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC ACC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCG TAG GGG ACC CGA GGG GAC TCG TCG AGC

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CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CCG AGC AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC AAC TCT ACC TTC
CCG GAC GTC GGG TGG GTC CCA CGG TAC GCC CGG AAG TTG AGA TGG AAG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCC TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGC CAG AAT TCC GTG GAA CGG GTC CGG

k) SEQ ID NO:12

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CGG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC GTT AAC GCT ACC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCC TAG CAA TTG CGA TGG GAC TCG TCG ACG

CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CCG AGC AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CCG GAC GTC GGG TGG GTC CCA CGG TAC GCC CGG AAG CGG AGA CGA AAG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCC TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGC CAG AAT TCC GTG GAA CGG GTC CGG

1) SEQ ID NO:13

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

- 13 -

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC CTC CCTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCG TAG GGG ACC CGA GGG GAC TCG TCG AGC

CCC AGC AAC GCC ACC CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG TTG CGG TGG GTC GAC CGT CGG AGC AAC TCG TTG GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC CTG GGC TCC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG

CGC CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC GCG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC KTC XAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC GGG

m) SEQ ID NO:14
ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCC GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC CTC CCTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCG TAG GGG ACC CGA GGG GAC TCG TCG AGC

CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CGG AGC AAC TCG TTG GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

AAC GGT ACC GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
TTG CCA TGG CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC

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CGG GAC GTC GGG TGG GTC CCA CGG TAC GCC CGG AAG CGG AGA CGA AAG
CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG
CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGC CAG AAT TCC GTG GAA CGG GTC CGG

n) SEQ ID NO:15

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA ATA TTG TGG TGG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC GTT AAC GCT ACC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCG TAG CAA TTG GCA TGG GAC TCG TCG AGC

CCC AGC AAC GCC ACC CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG TTG CGG TGG GTC GAC CGT CCG AGC AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CCG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGC CAG AAT TCC GTG GAA CGG GTC CGG

o) SEQ ID NO:16

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA ATA TTG TGG TGG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AAT TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TTA ACC

ACC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
TGG TCG GTC CGG GAC GTC GAC CGT CCG AGC AAC TCG GTT GAG GTA TCG

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GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG AAC GGG ACC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC TTG CCC TGG AGG
CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG
TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CCT CTT GAC CCT TAC CGG GGA
GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG
CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGC GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG
CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGC CAG AAT TCC GTG GAA CGG GTC GGG

A hyperglycosylated heterologous fusion protein of the present invention also includes polynucleotides encoding the heterologous fusion protein described herein, vectors comprising these polynucleotides and host cells transfected or transformed with the vectors described herein. Also included is a process for producing a heterologous fusion protein comprising the steps of transcribing and translating a polynucleotide described herein under conditions wherein the heterologous fusion protein is expressed in detectable amounts.

The present invention encompasses a method for increasing neutrophil levels in a mammal comprising the administration of a therapeutically effective amount of a heterologous fusion protein described above. The present invention also includes the use of the heterologous fusion proteins described above for the manufacture of a medicament for the treatment of patients with insufficient circulating neutrophil levels.

The present invention also encompasses a pharmaceutical formulation adapted for the treatment of patients with insufficient neutrophil levels comprising a glycosylated protein as described above.

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BRIEF DESCRIPTION OF THE FIGURES

The invention is further illustrated with reference to the following drawings:

Figure 1: Schematic illustrating fourteen regions in human G-CSF wherein the amino acid sequence can be mutated to create functional glycosylation sites.

Figure 2a: IgG1 Fc amino acid sequence encompassing the hinge region, CH2 and CH3 domains.

Figure 2b: IgG4 Fc amino acid sequence encompassing the hinge region, CH2 and CH3 domains.

Figure 3: Human serum albumin amino acid sequence

Figure 4: IgG1 Fc DNA sequence

Figure 5: IgG4 Fc DNA sequence with Ser229Pro mutation.

Figure 6: G-CSF/IgG1 Fc fusion protein

Figure 7: G-CSF/IgG4 Fc fusion protein

Figure 8: G-CSF/HA fusion protein.

The present invention comprises a heterologous fusion protein. As used herein, the term heterologous fusion protein means a hyperglycosylated G-CSF analog fused to human albumin, a human albumin analog, a human albumin fragment, the Fc portion of an immunoglobulin, an analog of the Fc portion of an immunoglobulin, or a fragment of the Fc portion of an immunoglobulin. The G-CSF analog may be fused directly, or fused via a peptide linker, to an albumin or Fc protein. The albumin and Fc portion may be fused to the G-CSF analogs at either terminus or at both termini. These heterologous fusion proteins are biologically active and have an increased half-life compared to native G-CSF.

Hyperglycosylated G-CSF Analogs

Encompassed by the invention are certain hyperglycosylated analogs of G-CSF. Analogs of G-CSF refer to human G-CSF with one or more changes in the amino acid

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sequence which result in an increase in the number of sites for carbohydrate attachment compared with native human G-CSF expressed in animal cells *in vivo*. In addition, G-CSF analogs include human G-CSF wherein the O-linked glycosylation site at position 133 is replaced with an N-linked glycosylation site. Analogs are generated by site directed mutagenesis having substitution of amino acid residues creating new sites that are available for glycosylation. Analogs having a greater carbohydrate content than that found in native human G-CSF are generated by adding glycosylation sites that do not perturb the secondary, tertiary, and quaternary structure required for activity. Furthermore, because the hyperglycosylated analogs of the present invention have a larger mass and an increased negative charge compared to native G-CSF, they will not be as rapidly cleared from the circulation.

It is preferred that the G-CSF analog have 1, 2, 3, or 4 additional sites for N-glycosylation. Figure 1 illustrates fourteen different regions that can be glycosylated with very little effect on *in vitro* activity. Each region may be mutated to the consensus site for N-glycosylation addition which is Asn X1 X2 wherein X1 is any amino acid except Pro and X2 is Ser or Thr. It is preferred that the X1 amino acid be any other amino acid except Trp, Asp, Glu, or Leu and it is most preferred that the X1 amino acid be the naturally occurring amino acid. The scope of the present invention includes analogs wherein a single region (1 through 14) is mutated or wherein a region is mutated in combination with one or more other regions.

Analogs having carbohydrate attached to only a single mutated site have been expressed, purified, characterized, and tested for activity. Similarly analogs with multiple glycosylation sites have been expressed, purified, characterized, and tested for activity. For example G-CSF[A37N,Y39T] is G-CSF wherein the amino acids at positions 37 and 39 have been substituted to create a glycosylation

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site. This site of carbohydrate attachment is illustrated as region 1 in Figure 1. G-CSF [A37N, Y39T, P57V, W58N, P60T] is an example of a G-CSF analog wherein amino acids in region 1 and region 2 are mutated to provide two functional glycosylation sites on a single molecule (Figure 1).

G-CSF [A37N, Y39T, P57V, W58N, P60T, Q67N, L69T] is an example of a G-CSF analog wherein the amino acids in region 1, region 2, and region 9 are mutated to provide three functional glycosylation sites on a single molecule (Figure 1).

Native G-CSF can be used as the backbone to create the glycosylated G-CSF analogs of the present invention. In addition, the native G-CSF backbone used to create the analogs of the present invention can be modified such that substitutions in the regions defined in Figure 1 are made in the context of a different or improved G-CSF protein. For example, native G-CSF with a Cysteine to Alanine substitution at position 17 may reduce aggregation and enhance stability and thus, can be used as the backbone used to create the glycosylated G-CSF analogs of the present invention.

In addition, Reidhaar-Olson et al., through alanine scanning mutagenesis, describe residues critical to the activity of human G-CSF. [Reidhaar-Olson et al. (1996) *Biochemistry* 35:9034-9041; See also Young et al. (1997) *Protein Science* 6:1228-1236]. Thus, the glycosylated analogs of the present invention can be modified by substituting amino acids outside the glycosylated regions described in Figure 1.

As outlined above, amino acid substitutions in the fusion proteins of the present invention can be based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity,

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hydrophilicity, charge, size, etc. Furthermore, substitutions can be made based on secondary structure propensity. For example, a helical amino acid can be replaced with an amino acid that would preserve the helical structure. Exemplary substitutions that take various of the foregoing characteristics into consideration in order to produce conservative amino acid changes resulting in silent changes within the present peptides, etc., can be selected from other members of the class to which the naturally occurring amino acid belongs.

The present invention also encompasses G-CSF analogs wherein the O-linked glycosylation site at position 133 is mutated to serve as an N-linked glycosylation site. The N-linked carbohydrate will generally have a higher sialic acid content which will protect it from the rapid clearance mechanisms associated with native G-CSF.

The functions of a carbohydrate chain greatly depends on the structure of the attached carbohydrate moiety. Typically compounds with a higher sialic acid content will have better stability and longer half-lives *in vivo*. The N-linked oligosaccharides contain sialic acid in both an α 2,3 and an α 2,6 linkage to galactose. [Takeuchi et al. (1988) *J. Biol. Chem.* 263:3657]. Typically the sialic acid in the α 2,3 linkage is added to galactose on the mannose α 1,6 branch and the sialic acid in the α 2,6 linkage is added to the galactose on the mannose α 1,3 branch. The enzymes that add these sialic acids (β -galactoside α 2,3 sialyltransferase and β -galactoside α 2,6 sialyltransferase) are most efficient at adding sialic acid to the mannose α 1,6 and mannose α 1,3 branches respectively.

Tetra-antennary N-linked oligosaccharides most commonly provide four possible sites for sialic acid attachment while bi- and tri-antennary oligosaccharide chains, which can substitute for the tetra-antennary form at Asn-like sites, commonly have at most only two or three sialic acids attached. O-linked oligosaccharides commonly provide only

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two sites for sialic acid attachment. Mammalian cell cultures can be screened for those cells that preferentially add teta-antennary chains to the G-CSF analogs of the present invention, thereby maximizing the number of sites for sialic acid attachment. Different types of mammalian cells also differ with respect to the transferase enzymes present and consequently the sialic acid content and type of oligosaccharide attached at each site. One way to optimize the carbohydrate content for a given G-CSF analog is to express the analog in a cell line wherein an expression plasmid containing DNA encoding a specific sialyl transferase (e.g., α 2,6 sialyltransferase) is co-transfected with the G-CSF analog expression plasmid. Alternatively a host cell line may be stably transfected with a sialyltransferase cDNA and that host cell used to express the G-CSF analog of interest. Thus, it is preferable if the oligosaccharide structure and sialic acid content are optimized for each analog encompassed by the present invention.

Heterologous Fc fusion proteins:

The hyperglycosylated G-CSF analogs described above can be fused directly or via a peptide linker to the Fc portion of an immunoglobulin. (See Figures 6-7).

Immunoglobulins are molecules containing polypeptide chains held together by disulfide bonds, typically having two light chains and two heavy chains. In each chain, one domain (V) has a variable amino acid sequence depending on the antibody specificity of the molecule. The other domains (C) have a rather constant sequence common to molecules of the same class.

As used herein, the Fc portion of an immunoglobulin has the meaning commonly given to the term in the field of immunology. Specifically, this term refers to an antibody fragment which is obtained by removing the two antigen binding regions (the Fab fragments) from the antibody.

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Thus, the Fc portion is formed from approximately equal sized fragments of the constant region from both heavy chains, which associate through non-covalent interactions and disulfide bonds. The Fc portion can include the hinge regions and extend through the CH₂ and CH₃ domains to the C-terminus of the antibody. Representative hinge regions for human and mouse immunoglobulins can be found in Antibody Engineering, A Practical Guide, Borrebaeck, C.A.K., ed., W.H. Freeman and Co., 1992, the teachings of which are herein incorporated by reference. The amino acid sequence of a representative Fc protein containing a hinge region, CH₂ and CH₃ domains is shown in Figures 2a and 2b.

There are five types of human immunoglobulin Fc regions with different effector and pharmacokinetic properties: IgG, IgA, IgM, IgD, and IgE. IgG is the most abundant immunoglobulin in serum. IgG also has the longest half-life in serum of any immunoglobulin (23 days). Unlike other immunoglobulins, IgG is efficiently recirculated following binding to an Fc receptor. There are four IgG subclasses G1, G2, G3, and G4, each of which have different effector functions. G1, G2, and G3 can bind Clq and fix complement while G4 cannot. Even though G3 is able to bind Clq more efficiently than G1, G1 is more effective at mediating complement-directed cell lysis. G2 fixes complement very inefficiently. The Clq binding site in IgG is located at the carboxy terminal region of the CH₂ domain.

All IgG subclasses are capable of binding to Fc receptors (CD16, CD32, CD64) with G1 and G3 being more effective than G2 and G4. The Fc receptor-binding region of IgG is formed by residues located in both the hinge and the carboxy terminal regions of the CH₂ domain.

IgA can exist both in a monomeric and dimeric form held together by a J-chain. IgA is the second most abundant Ig in serum, but it has a half-life of only 6 days. IgA has three effector functions. It binds to an IgA specific receptor on macrophages and eosinophils, which drives

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phagocytosis and degranulation, respectively. It can also fix complement via an unknown alternative pathway.

IgM is expressed as either a pentamer or a hexamer, both of which are held together by a J-chain. IgM has a serum half-life of 5 days. It binds weakly to C1q via a binding site located in its CH3 domain. IgD has a half-life of 3 days in serum. It is unclear what effector functions are attributable to this Ig. IgE is a monomeric Ig and has a serum half-life of 2.5 days. IgE binds to two Fc receptors which drives degranulation and results in the release of proinflammatory agents.

Depending on the desired *in vivo* effect, the heterologous fusion proteins of the present invention may contain any of the isotypes described above or may contain mutated Fc regions wherein the complement and/or Fc receptor binding functions have been altered. For example, one embodiment of the present invention is a Ser229Pro mutation in IgG4 Fc, which reduces monomer formation. See Figure 5.

The heterologous fusion proteins of the present invention may contain the entire Fc portion of an immunoglobulin, fragments of the Fc portion of an immunoglobulin, or analogs thereof fused to a G-CSF analog. Furthermore, the Fc portion may be fused at either terminus or at both termini.

The heterologous fusion proteins of the present invention can consist of single chain proteins or as multi-chain polypeptides. Two or more Fc fusion proteins can be produced such that they interact through disulfide bonds that naturally form between Fc regions. These multimers can be homogeneous with respect to the G-CSF analog or they may contain different G-CSF analogs fused at the N-terminus of the Fc portion of the fusion protein.

Regardless of the final structure of the fusion protein, the Fc or Fc-like region must serve to prolong the *in vivo* plasma half-life of the G-CSF analog compared to the native G-CSF. Furthermore, the fused G-CSF analog must

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retain some biological activity. Biological activity can be determined by *in vitro* and *in vivo* methods known in the art.

Since the Fc region of IgG produced by proteolysis has the same *in vivo* half-life as the intact IgG molecule and Fab fragments are rapidly degraded, it is believed that the relevant sequence for prolonging half-life resides in the CH₂ and/or CH₃ domains. Further, it has been shown in the literature that the catabolic rates of IgG variants that do not bind the high-affinity Fc receptor or C1q are indistinguishable from the rate of clearance of the parent wild-type antibody, indicating that the catabolic site is distinct from the sites involved in Fc receptor or C1q binding. [Wawrzynczak et al., (1992) *Molecular Immunology* 29:221]. Site-directed mutagenesis studies using a murine IgG1 Fc region suggested that the site of the IgG1 Fc region that controls the catabolic rate is located at the CH₂-CH₃ domain interface.

Based on these studies, Fc regions can be modified at the catabolic site to optimize the half-life of the fusion proteins. It is preferable that the Fc region used for the heterologous fusion proteins of the present invention be derived from an IgG1 (see Figure 4) or an IgG4 Fc region. It is even more preferable that the Fc region be IgG4 or derived from IgG4. Preferably the IgG Fc region contains both the CH₂ and CH₃ regions including the hinge region.

Heterologous albumin fusion proteins:

The G-CSF analogs described above can be fused directly or via a peptide linker to albumin or an analog, fragment, or derivative thereof. (See Figure 8).

Generally the albumin proteins making up part of the fusion proteins of the present invention can be derived from albumin cloned from any species. However, human albumin and fragments and analogs thereof are preferred to reduce the risk of the fusion protein being immunogenic in humans. Human serum albumin (HA) consists of a single non-

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glycosylated polypeptide chain of 585 amino acids with a formula molecular weight of 66,500. The amino acid sequence of human HA is shown in Figure 3. [See Meloun, et al. (1975) FEBS Letters 58:136; Behrens, et al. (1975) Fed. Proc. 34:591; Lawn, et al. (1981) Nucleic Acids Research 9:6102-6114; Minghetti, et al. (1986) J. Biol. Chem. 261:6747]. A variety of polymorphic variants as well as analogs and fragments of albumin have been described. [See Weitkamp, et al., (1973) Ann. Hum. Genet. 37:219]. For example, in EP 322,094, the inventors disclose various shorter forms of HA. Some of these fragments include HA(1-373), HA(1-388), HA(1-389), HA(1-369), and HA(1-419) and fragments between 1-369 and 1-419. EP 399,666 discloses albumin fragments that include HA(1-177) and HA(1-200) and fragments between HA(1-177) and HA(1-200).

It is understood that the heterologous fusion proteins of the present invention include G-CSF analogs that are coupled to any albumin protein including fragments, analogs, and derivatives wherein such fusion protein is biologically active and has a longer plasma half-life than the G-CSF analog alone. Thus, the albumin portion of the fusion protein need not necessarily have a plasma half-life equal to that of native human albumin. In addition, the albumin may be fused to either terminus or both termini of the hyperglycosylated G-CSF analog. Fragments, analogs, and derivatives are known or can be generated that have longer half-lives or have half-lives intermediate to that of native human albumin and the G-CSF analog of interest.

The heterologous fusion proteins of the present invention encompass proteins having conservative amino acid substitutions in the G-CSF analog and/or the Fc or albumin portion of the fusion protein. A "conservative substitution" is the replacement of an amino acid with another amino acid that has the same net electronic charge and approximately the same size and shape. Amino acids with aliphatic or substituted aliphatic amino acid side chains

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have approximately the same size when the total number carbon and heteroatoms in their side chains differs by no more than about four. They have approximately the same shape when the number of branches in their side chains differs by no more than one. Amino acids with phenyl or substituted phenyl groups in their side chains are considered to have about the same size and shape. Except as otherwise specifically provided herein, conservative substitutions are preferably made with naturally occurring amino acids.

However, the term "amino acid" is used herein in its broadest sense, and includes naturally occurring amino acids as well as non-naturally occurring amino acids, including amino acid analogs and derivatives. The latter includes molecules containing an amino acid moiety. One skilled in the art will recognize, in view of this broad definition, that reference herein to an amino acid includes, for example, naturally occurring proteogenic L-amino acids; D-amino acids; chemically modified amino acids such as amino acid analogs and derivatives; naturally occurring non-proteogenic amino acids such as norleucine, β -alanine, ornithine, GABA, etc.; and chemically synthesized compounds having properties known in the art to be characteristic of amino acids. As used herein, the term "proteogenic" indicates that the amino acid can be incorporated into a peptide, polypeptide, or protein in a cell through a metabolic pathway.

The incorporation of non-natural amino acids, including synthetic non-native amino acids, substituted amino acids, or one or more D-amino acids into the heterologous fusion proteins of the present invention can be advantageous in a number of different ways. D-amino acid-containing peptides, etc., exhibit increased stability *in vitro* or *in vivo* compared to L-amino acid-containing counterparts. Thus, the construction of peptides, etc., incorporating D-amino acids can be particularly useful when greater intracellular

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stability is desired or required. More specifically, D-peptides, etc., are resistant to endogenous peptidases and proteases, thereby providing improved bioavailability of the molecule, and prolonged lifetimes *in vivo* when such properties are desirable. Additionally, D-peptides, etc., cannot be processed efficiently for major histocompatibility complex class II-restricted presentation to T helper cells, and are therefore less likely to induce humoral immune responses in the whole organism.

General methods for making the heterologous fusion proteins of the present invention.

Although the heterologous fusion proteins of the present invention can be made by a variety of different methods, recombinant methods are preferred. For purposes of the present invention, as disclosed and claimed herein, the following general molecular biology terms and abbreviations are defined below. The terms and abbreviations used in this document have their normal meanings unless otherwise designated. For example, "°C" refers to degrees Celsius; "mmol" refers to millimole or millimoles; "mg" refers to milligrams; "μg" refers to micrograms; "ml or mL" refers to milliliters; and "μl or μL" refers to microliters. Amino acids abbreviations are as set forth in 37 C.F.R. § 1.822 (b) (2) (1994).

"Base pair" or "bp" as used herein refers to DNA or RNA. The abbreviations A,C,G, and T correspond to the 5'-monophosphate forms of the deoxyribonucleosides (deoxy)adenosine, (deoxy)cytidine, (deoxy)guanosine, and thymidine, respectively, when they occur in DNA molecules. The abbreviations U,C,G, and A correspond to the 5'-monophosphate forms of the ribonucleosides uridine, cytidine, guanosine, and adenosine, respectively when they occur in RNA molecules. In double stranded DNA, base pair may refer to a partnership of A with T or C with G. In a DNA/RNA, heteroduplex base pair may refer to a partnership

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of A with U or C with G. (See the definition of "complementary", *infra*.)

"Digestion" or "Restriction" of DNA refers to the catalytic cleavage of the DNA with a restriction enzyme that acts only at certain sequences in the DNA ("sequence-specific endonucleases"). The various restriction enzymes used herein are commercially available and their reaction conditions, cofactors, and other requirements were used as would be known to one of ordinary skill in the art. Appropriate buffers and substrate amounts for particular restriction enzymes are specified by the manufacturer or can be readily found in the literature.

"Ligation" refers to the process of forming phosphodiester bonds between two double stranded nucleic acid fragments. Unless otherwise provided, ligation may be accomplished using known buffers and conditions with a DNA ligase, such as T4 DNA ligase.

"Plasmid" refers to an extrachromosomal (usually) self-replicating genetic element. Plasmids are generally designated by a lower case "p" followed by letters and/or numbers. The starting plasmids herein are either commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids in accordance with published procedures. In addition, equivalent plasmids to those described are known in the art and will be apparent to the ordinarily skilled artisan.

"Recombinant DNA cloning vector" as used herein refers to any autonomously replicating agent, including, but not limited to, plasmids and phages, comprising a DNA molecule to which one or more additional DNA segments can or have been added.

"Recombinant DNA expression vector" as used herein refers to any recombinant DNA cloning vector in which a promoter to control transcription of the inserted DNA has been incorporated.

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"Transcription" refers to the process whereby information contained in a nucleotide sequence of DNA is transferred to a complementary RNA sequence.

"Transfection" refers to the uptake of an expression vector by a host cell whether or not any coding sequences are, in fact, expressed. Numerous methods of transfection are known to the ordinarily skilled artisan, for example, calcium phosphate co-precipitation, liposome transfection, and electroporation. Successful transfection is generally recognized when any indication of the operation of this vector occurs within the host cell.

"Transformation" refers to the introduction of DNA into an organism so that the DNA is replicable, either as an extrachromosomal element or by chromosomal integration. Methods of transforming bacterial and eukaryotic hosts are well known in the art, many of which methods, such as nuclear injection, protoplast fusion or by calcium treatment using calcium chloride are summarized in J. Sambrook, et al., *Molecular Cloning: A Laboratory Manual*, (1989). Generally, when introducing DNA into Yeast the term transformation is used as opposed to the term transfection.

"Translation" as used herein refers to the process whereby the genetic information of messenger RNA (mRNA) is used to specify and direct the synthesis of a polypeptide chain.

"Vector" refers to a nucleic acid compound used for the transfection and/or transformation of cells in gene manipulation bearing polynucleotide sequences corresponding to appropriate protein molecules which, when combined with appropriate control sequences, confers specific properties on the host cell to be transfected and/or transformed. Plasmids, viruses, and bacteriophage are suitable vectors. Artificial vectors are constructed by cutting and joining DNA molecules from different sources using restriction enzymes and ligases. The term "vector" as used herein

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includes Recombinant DNA cloning vectors and Recombinant DNA expression vectors.

"Complementary" or "Complementarity", as used herein, refers to pairs of bases (purines and pyrimidines) that associate through hydrogen bonding in a double stranded nucleic acid. The following base pairs are complementary: guanine and cytosine; adenine and thymine; and adenine and uracil.

"Hybridization" as used herein refers to a process in which a strand of nucleic acid joins with a complementary strand through base pairing. The conditions employed in the hybridization of two non-identical, but very similar, complementary nucleic acids varies with the degree of complementarity of the two strands and the length of the strands. Such techniques and conditions are well known to practitioners in this field.

"Isolated amino acid sequence" refers to any amino acid sequence, however, constructed or synthesized, which is locationally distinct from the naturally occurring sequence.

"Isolated DNA compound" refers to any DNA sequence, however constructed or synthesized, which is locationally distinct from its natural location in genomic DNA.

"Isolated nucleic acid compound" refers to any RNA or DNA sequence, however constructed or synthesized, which is locationally distinct from its natural location.

"Primer" refers to a nucleic acid fragment which functions as an initiating substrate for enzymatic or synthetic elongation.

"Promoter" refers to a DNA sequence which directs transcription of DNA to RNA.

"Probe" refers to a nucleic acid compound or a fragment, thereof, which hybridizes with another nucleic acid compound.

"Stringency" of hybridization reactions is readily determinable by one of ordinary skill in the art, and generally is an empirical calculation dependent upon probe

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length, washing temperature, and salt concentration. In general, longer probes require higher temperatures for proper annealing, while short probes need lower temperatures. Hybridization generally depends on the ability of denatured DNA to re-anneal when complementary strands are present in an environment below their melting temperature. The higher the degree of desired homology between the probe and hybridizable sequence, the higher the relative temperature that can be used. As a result, it follows that higher relative temperatures would tend to make the reactions more stringent, while lower temperatures less so. For additional details and explanation of stringency of hybridization reactions, see Ausubel et al., *Current Protocols in Molecular Biology*, Wiley Interscience Publishers, 1995.

"Stringent conditions" or "high stringency conditions", as defined herein, may be identified by those that (1) employ low ionic strength and high temperature for washing, for example, 15 mM sodium chloride/1.5 mM sodium citrate/0.1% sodium dodecyl sulfate at 50°C; (2) employ during hybridization a denaturing agent, such as formamide, for example, 50% (v/v) formamide with 0.1% bovine serum albumin/0.1% ficoll/0.1% polyvinylpyrrolidone/50 mM sodium phosphate buffer at pH 6.5 with 750 mM sodium chloride/75 mM sodium citrate at 42°C; or (3) employ 50% formamide, 5X SSC (750 mM sodium chloride, 75 mM sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5X Denhardt's solution, sonicated salmon sperm DNA (50 µg/ml), 0.1% SDS, and 10% dextran sulfate at 42°C with washes at 42°C in 0.2X SSC (30 mM sodium chloride/3 mM sodium citrate) and 50% formamide at 55°C, followed by a high-stringency wash consisting of 0.1X SSC containing EDTA at 55°C.

"Moderately stringent conditions" may be identified as described by Sambrook et al. [*Molecular Cloning: A Laboratory Manual*, New York: Cold Spring Harbor Press, (1989)], and include the use of washing solution and hybridization conditions (e.g., temperature, ionic strength, and %SDS) less

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stringent than those described above. An example of moderately stringent conditions is overnight incubation at 37°C in a solution comprising: 20% formamide, 5X SSC (750 mM sodium chloride, 75 mM sodium citrate), 50 mM sodium phosphate at pH 7.6, 5X Denhardt's solution, 10% dextran sulfate, and 20 mg/mL denatured sheared salmon sperm DNA, followed by washing the filters in 1X SSC at about 37-50°C. The skilled artisan will recognize how to adjust the temperature, ionic strength, etc., as necessary to accommodate factors such as probe length and the like.

"PCR" refers to the widely-known polymerase chain reaction employing a thermally-stable DNA polymerase.

"Leader sequence" refers to a sequence of amino acids which can be enzymatically or chemically removed to produce the desired polypeptide of interest.

"Secretion signal sequence" refers to a sequence of amino acids generally present at the N-terminal region of a larger polypeptide functioning to initiate association of that polypeptide with the cell membrane and secretion of that polypeptide through the cell membrane.

Construction of DNA encoding the heterologous fusion proteins of the present invention:

Wild type albumin and immunoglobulin proteins can be obtained from a variety of sources. For example, these proteins can be obtained from a cDNA library prepared from tissue or cells which express the mRNA of interest at a detectable level. Libraries can be screened with probes designed using the published DNA or protein sequence for the particular protein of interest.

Screening a cDNA or genomic library with the selected probe may be conducted using standard procedures, such as described in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, NY (1989). An alternative means to isolate a gene encoding an albumin or immunoglobulin protein is to use PCR methodology

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[Sambrook et al., *supra*; Dieffenbach et al., *PCR Primer: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, NY (1995)]. PCR primers can be designed based on published sequences.

Generally the full-length wild-type sequences cloned from a particular species can serve as a template to create analogs, fragments, and derivatives that retain the ability to confer a longer plasma half-life on the G-CSF analog that is part of the fusion protein. It is preferred that the Fc and albumin portions of the heterologous fusion proteins of the present invention be derived from the native human sequence in order to reduce the risk of potential immunogenicity of the fusion protein in humans.

In particular, it is preferred that the immunoglobulin portion of a fusion protein encompassed by the present invention contain only an Fc fragment of the immunoglobulin. Depending on whether particular effector functions are desired and the structural characteristics of the fusion protein, an Fc fragment may contain the hinge region along with the CH₂ and CH₃ domains or some other combination thereof. These Fc fragments can be generated using PCR techniques with primers designed to hybridize to sequences corresponding to the desired ends of the fragment. Similarly, if fragments of albumin are desired, PCR primers can be designed which are complementary to internal albumin sequences. PCR primers can also be designed to create restriction enzyme sites to facilitate cloning into expression vectors.

DNA encoding human G-CSF can be obtained from a cDNA library prepared from tissue or cells which express G-CSF mRNA at a detectable level such as monocytes, macrophages, vascular endothelial cells, fibroblasts, and some human malignant and leukemic myeloblastic cells. Libraries can be screened with probes designed using the published DNA sequence for human G-CSF. [Souza L. et al. (1986) *Science* 232:61-65]. Screening a cDNA or genomic library with the

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selected probe may be conducted using standard procedures, such as described in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, NY (1989). An alternative means to isolate the gene encoding human G-CSF is to use PCR methodology [Sambrook et al., supra; Dieffenbach et al., *PCR Primer: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, NY (1995)].

The glycosylated G-CSF analogs of the present invention can be constructed by a variety of mutagenesis techniques well known in the art. Specifically, a representative number of glycosylated G-CSF analogs were constructed using mutagenic PCR from a cloned wild-type human G-CSF DNA template (Example 1).

The glycosylated G-CSF analogs of the present invention may be produced by other methods including recombinant DNA technology or well known chemical procedures, such as solution or solid-phase peptide synthesis, or semi-synthesis in solution beginning with protein fragments coupled through conventional solution methods.

Recombinant DNA methods are preferred for producing the glycosylated G-CSF analogs of the present invention. Host cells are transfected or transformed with expression or cloning vectors described herein for glycosylated G-CSF analog production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences (Example 2). The culture conditions, such as media, temperature, pH and the like, can be selected by the skilled artisan without undue experimentation.

Physical stability is an essential feature for therapeutic formulations. The physical stability of the heterologous fusion proteins of the present invention depends on their conformational stability, the number of charged residues (*pI* of the protein), the ionic strength and pH of the formulation, and the protein concentration, among other possible factors. As discussed previously, the G-CSF

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analog portion of the heterologous fusion proteins can be successfully glycosylated and expressed such that they maintain their three dimensional structure. Because these analogs are able to fold properly in a hyperglycosylated state, they will have improved conformational and physical stability relative to wild-type G-CSF.

While wild-type G-CSF produced in mammalian cells and bacterial cells has similar activity *in vivo*, the mammalian cell-produced protein has increased conformational and physical stability due to the presence of a single O-linked sugar moiety present at position 133. Thus, the G-CSF analog portion of the heterologous fusion proteins, which have an increased glycosylation content compared to wild-type G-CSF produced in mammalian or bacterial cells, will have increased stability. Furthermore, it is likely that glycosylation may inhibit inter-domain interactions and consequently enhance stability by preventing inter-domain disulfide shuffling.

The gene encoding a heterologous fusion protein can be constructed by ligating DNA encoding a G-CSF analog in-frame to DNA encoding an albumin or Fc protein. The gene encoding the G-CSF analog and the gene encoding the albumin or Fc protein can also be joined in-frame via DNA encoding a linker peptide.

The *in vivo* function and stability of the heterologous fusion proteins of the present invention can be optimized by adding small peptide linkers to prevent potentially unwanted domain interactions. Although these linkers can potentially be any length and consist of any combination of amino acids, it is preferred that the length be no longer than necessary to prevent unwanted domain interactions and/or optimize biological activity and/or stability. Generally, the linkers should not contain amino acids with extremely bulky side chains or amino acids likely to introduce significant secondary structure. It is preferred that the linker be serine-glycine rich and be less than 30 amino acids in

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length. It is more preferred that the linker be no more than 20 amino acids in length. It is even more preferred that the linker be no more than 15 amino acids in length. A preferred linker contains repeats of the sequence Gly-Gly-Gly-Gly-Ser. It is preferred that there be between 2 and 6 repeats of this sequence. It is even more preferred that there be between 3 and 4 repeats of this sequence.

To construct the heterologous G-CSF fusion proteins, the DNA encoding wild-type G-CSF, albumin, and Fc polypeptides and fragments thereof can be mutated either before ligation or in the context of a cDNA encoding an entire fusion protein. A variety of mutagenesis techniques are well known in the art. For example, a mutagenic PCR method utilizes strand overlap extension to create specific base mutations for the purposes of changing a specific amino acid sequence in the corresponding protein. This PCR mutagenesis requires the use of four primers, two in the forward orientation (primers A and C) and two in the reverse orientation (primers B and D). A mutated gene is amplified from the wild-type template in two different stages. The first reaction amplifies the gene in halves by performing an A to B reaction and a separate C to D reaction wherein the B and C primers target the area of the gene to be mutated. When aligning these primers with the target area, they contain mismatches for the bases that are targeted to be changed. Once the A to B and C to D reactions are complete, the reaction products are isolated and mixed for use as the template for the A to D reaction. This reaction then yields the full, mutated product.

Once a gene encoding an entire fusion protein is produced it can be cloned into an appropriate expression vector. Specific strategies that can be employed to make the G-CSF fusion proteins of the present invention are described in example 1.

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General methods to recombinantly express the heterologous fusion proteins of the present invention:

Host cells are transfected or transformed with expression or cloning vectors described herein for heterologous fusion protein production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences. The culture conditions, such as media, temperature, pH and the like, can be selected by the skilled artisan without undue experimentation. In general, principles, protocols, and practical techniques for maximizing the productivity of cell cultures can be found in *Mammalian Cell Biotechnology: A Practical Approach*, M. Butler, ed. (IRL Press, 1991) and Sambrook, et al., *supra*. Methods of transfection are known to the ordinarily skilled artisan, for example, CaPO₄ and electroporation. General aspects of mammalian cell host system transformations have been described in U.S. Patent No. 4,399,216. Transformations into yeast are typically carried out according to the method of van Solingen et al., *J Bact.* 130(2): 946-7 (1977) and Hsiao et al., *Proc. Natl. Acad. Sci. USA* 76(8): 3829-33 (1979). Suitable host cells for the expression of the fusion proteins of the present invention are derived from multicellular organisms.

The fusion proteins of the present invention may be recombinantly produced directly, or as a protein having a signal sequence or other additional sequences which create a specific cleavage site at the N-terminus of the mature fusion protein. In general, the signal sequence may be a component of the vector, or it may be a part of the fusion protein-encoding DNA that is inserted into the vector. The signal sequence may be a prokaryotic signal sequence selected, for example, from the group of the alkaline phosphatase, penicillinase, lpp, or heat-stable enterotoxin II leaders. For yeast secretion the signal sequence may be, e.g., the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces* cc-factor leaders, the latter

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described in U.S. Patent No. 5,010,182), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179), or the signal described in WO 90/13646. In mammalian cell expression, mammalian signal sequences may be used to direct secretion of the protein, such as signal sequences from secreted polypeptides of the same or related species as well as viral secretory leaders.

Both expression and cloning vectors contain a nucleic acid sequence that enables the vector to replicate in one or more selected host cells. Such sequences are well known for a variety of bacteria, yeast, and viruses. The origin of replication from the plasmid pBR322 is suitable for most Gram-negative bacteria, the 2u plasmid origin is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells. Expression and cloning vectors will typically contain a selection gene, also termed a selectable marker. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, e.g., ampicillin, neomycin, methotrexate, or tetracycline, (b) complement autotrophic deficiencies, or (c) supply critical nutrients not available from complex media, e.g., the gene encoding D-alanine racemase for *Bacilli*.

An example of suitable selectable markers for mammalian cells are those that enable the identification of cells competent to take up the fusion protein-encoding nucleic acid, such as DHFR or thymidine kinase. An appropriate host cell when wild-type DHFR is employed is the CHO cell line deficient in DHFR activity, prepared and propagated as described [Urlaub and Chasin, *Proc. Natl. Acad. Sci. USA*, 77(7): 4216-20 (1980)]. A suitable selection gene for use in yeast is the *trpl* gene present in the yeast plasmid Yrp7 [Stinchcomb, et al., *Nature* 282(5734): 39-43 (1979); Kingsman, et al., *Gene* 7(2): 141-52 (1979); Tschumper, et al., *Gene* 10(2): 157-66 (1980)]. The *trpl* gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in

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tryptophan, for example, ATCC No. 44076 or PEPC1 [Jones, *Genetics* 85: 23-33 (1977)].

Expression and cloning vectors usually contain a promoter operably linked to the fusion protein-encoding nucleic acid sequence to direct mRNA synthesis. Promoters recognized by a variety of potential host cells are well known. Promoters suitable for use with prokaryotic hosts include the β -lactamase and lactose promoter systems [Chang, et al., *Nature* 275(5681): 617-24 (1978); Goeddel, et al., *Nature* 281(5732): 544-8 (1979)], alkaline phosphatase, a tryptophan (up) promoter system [Goeddel, *Nucleic Acids Res.* 8(18): 4057-74 (1980); EP 36,776 published 30 September 1981], and hybrid promoters such as the tat promoter [deBoer, et al., *Proc. Natl. Acad. Sci. USA* 80(1): 21-5 (1983)]. Promoters for use in bacterial systems also will contain a Shine-Dalgarno (S.D.) sequence operably linked to the DNA encoding the fusion protein.

Transcription of a polynucleotide encoding a fusion protein by higher eukaryotes may be increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp, that act on a promoter to increase its transcription. Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α -ketoprotein, and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. The enhancer may be spliced into the vector at a position 5' or 3' to the fusion protein coding sequence but is preferably located at a site 5' from the promoter.

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human, or nucleated cells from other multicellular organisms) will also contain sequences necessary for the termination of transcription and for

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stabilizing the mRNA. Such sequences are commonly available from the 5' and occasionally 3' untranslated regions of eukaryotic or viral DNAs or cDNAs. These regions contain nucleotide segments transcribed as polyadenylated fragments in the untranslated portion of the mRNA encoding the fusion protein.

Various forms of a fusion protein may be recovered from culture medium or from host cell lysates. If membrane-bound, it can be released from the membrane using a suitable detergent solution (e.g., Triton-X 100) or by enzymatic cleavage. Cells employed in expression of a fusion protein can be disrupted by various physical or chemical means, such as freeze-thaw cycling, sonication, mechanical disruption, or cell lysing agents.

Purification of the heterologous fusion proteins of the present invention:

Once the heterologous fusion proteins of the present invention are expressed in the appropriate host cell, the analogs can be isolated and purified. The following procedures are exemplary of suitable purification procedures:

Various methods of protein purification may be employed and such methods are known in the art and described, for example, in Deutscher, *Methods in Enzymology* 182: 83-9 (1990) and Scopes, *Protein Purification: Principles and Practice*, Springer-Verlag, NY (1982). The purification step(s) selected will depend on the nature of the production process used and the particular fusion protein produced. For example, fusion proteins comprising an Fc fragment can be effectively purified using a Protein A or Protein G affinity matrix. Low or high pH buffers can be used to elute the fusion protein from the affinity matrix. Mild elution conditions will aid in preventing irreversible denaturation of the fusion protein. Imidazole-containing buffers can also be used. Example 3 describes some successful purification protocols for the fusion proteins of the present invention.

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Characterization of the heterologous fusion proteins of the present invention:

Numerous methods exist to characterize the fusion proteins of the present invention. Some of these methods include: SDS-PAGE coupled with protein staining methods or immunoblotting using anti-IgG, anti-HA and anti-G-CSF antibodies. Other methods include matrix assisted laser desorption/ionization-mass spectrometry (MALDI-MS), liquid chromatography/mass spectrometry, isoelectric focusing, analytical anion exchange, chromatofocussing, and circular dichroism to name a few. A representative number of heterologous fusion proteins were characterized using SDS-PAGE coupled with immunoblotting as well as mass spectrometry.

For example, Table 2 illustrates the calculated molecular mass for a representative number of fusion proteins as well as the observed mass (as measured by protease mapping/LC-MS). The relative differences between observed mass and mass calculated for a nonglycosylated protein are indicative of the extent of glycosylation.

The heterologous fusion proteins of the present invention may be formulated with one or more excipients. The active fusion proteins of the present invention may be combined with a pharmaceutically acceptable buffer, and the pH adjusted to provide acceptable stability, and a pH acceptable for administration such as parenteral administration.

Optionally, one or more pharmaceutically-acceptable antimicrobial agents may be added. Meta-cresol and phenol are preferred pharmaceutically-acceptable microbial agents. One or more pharmaceutically-acceptable salts may be added to adjust the ionic strength or tonicity. One or more excipients may be added to adjust the isotonicity of the formulation. Glycerin is an example of an isotonicity-adjusting excipient. Pharmaceutically acceptable means suitable for administration to a human or other animal and thus, does not contain toxic

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elements or undesirable contaminants and does not interfere with the activity of the active compounds therein.

A pharmaceutically-acceptable salt form of the heterologous fusion proteins of the present invention may be used in the present invention. Acids commonly employed to form acid addition salts are inorganic acids such as hydrochloric acid, hydrobromic acid, hydriodic acid, sulfuric acid, phosphoric acid, and the like, and organic acids such as p-toluenesulfonic acid, methanesulfonic acid, oxalic acid, p-bromophenyl-sulfonic acid, carbonic acid, succinic acid, citric acid, benzoic acid, acetic acid, and the like. Preferred acid addition salts are those formed with mineral acids such as hydrochloric acid and hydrobromic acid.

Base addition salts include those derived from inorganic bases, such as ammonium or alkali or alkaline earth metal hydroxides, carbonates, bicarbonates, and the like. Such bases useful in preparing the salts of this invention thus include sodium hydroxide, potassium hydroxide, ammonium hydroxide, potassium carbonate, and the like.

Administration of Compositions:

Administration may be via any route known to be effective by the physician of ordinary skill. Peripheral, parenteral is one such method. Parenteral administration is commonly understood in the medical literature as the injection of a dosage form into the body by a sterile syringe or some other mechanical device such as an infusion pump. Peripheral parenteral routes can include intravenous, intramuscular, subcutaneous, and intraperitoneal routes of administration.

The heterologous fusion proteins of the present invention may also be amenable to administration by oral, rectal, nasal, or lower respiratory routes, which are non-parenteral routes. Of these non-parenteral routes, the lower respiratory route and the oral route are preferred.

The heterologous fusion proteins of the present invention can be used to treat patients with insufficient

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circulating neutrophil levels, typically those undergoing cancer chemotherapy.

An "effective amount" of the heterologous fusion protein is the quantity which results in a desired therapeutic and/or prophylactic effect without causing unacceptable side-effects when administered to a subject in need of G-CSF receptor stimulation. A "desired therapeutic effect" includes one or more of the following: 1) an amelioration of the symptom(s) associated with the disease or condition; 2) a delay in the onset of symptoms associated with the disease or condition; 3) increased longevity compared with the absence of the treatment; and 4) greater quality of life compared with the absence of the treatment.

The present invention comprises G-CSF compounds that have improved biochemical and biophysical properties by virtue of being fused to an albumin protein, an albumin fragment, an albumin analog, a Fc protein, a Fc fragment, or a Fc analog. These heterologous proteins can be successfully expressed in host cells, retain signaling activities associated with activation of the G-CSF receptor, and have prolonged half-lives.

The following examples are presented to further describe the present invention. The scope of the present invention is not to be construed as merely consisting of the following examples. Those skilled in the art will recognize that the particular reagents, equipment, and procedures described are merely illustrative and are not intended to limit the present invention in any manner.

EXAMPLES

Example 1: Construction of DNA encoding glycosylated G-CSF analogs:

Table 1 provides the sequence of primers used to create functional glycosylation sites in different regions of the protein (See Figure 1).

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Table 1: Primer sequences used to introduce mutations into human G-CSF.

Mutation	A Primer*	B Primer*	C Primer*	D Primer*
WT	CF177 [SEQ ID NO:25] GTAAGCTTGC GT CGACGCTAGCGG CGCGCCGCCATG GCCGGACCTGCC ACCCAGAGCCCC ATGAAGCTG	CF178 [SEQ ID NO:26] GGGGCAGGGAGC TGGCTGGGCCA GTGGAGTGGCTT CCTGCACTGTCC AGAGTGCACTGT G	CF179 [SEQ ID NO:27] GGACAGTGCAGG AAAGCCACTCCAC TGGGCCAGCCA GCTCCCTGCCCA AGAGCTTCCTG G	CF176 [SEQ ID NO:28] GAACCTCGAGGA TCCTCATTTAGGG CTGGGCAAGGTG CCTTAAGACGCG GTACGACACCTC CAGGAAGCTCTG
C17A <i>SacI</i>	CF177 [SEQ ID NO:29] GTAAGCTTGC GT CGACGCTAGCGG CGCGCCGCCATG GCCGGACCTGCC ACCCAGAGCCCC ATGAAGCTG	C17Arev [SEQ ID NO:30] GCTCTAACGGCCT TGAGCAGGAAGC TCTGGGGCAGGG AGCTCGCTGGGC CCAGTGGAG	C17Af or [SEQ ID NO:31] GGGCCAGCGAG CTCCCTGCCCA GAGCTTCCTGCT CAAGGGCTTAGA GCAAG	CF176 [SEQ ID NO:32] GAACCTCGAGGA TCCTCATTTAGGG CTGGGCAAGGTG CCTTAAGACGCG GTACGACACCTC CAGGAAGCTCTG
A37N, Y39T <i>SpeI</i>	CF177 [SEQ ID NO:33] GTAAGCTTGC GT CGACGCTAGCGG CGCGCCGCCATG GCCGGACCTGCC ACCCAGAGCCCC ATGAAGCTG	A37Nrev [SEQ ID NO:34] GTCGGAGCAGCA CTAGTCCCTCGG GGTGGCACAGCT TGGTGGTGT TAC ACAGCTTCCT G	A37Nfor [SEQ ID NO:35] GGCGCAGCGCTC CAGGAGAACGCTG TGTAAACACCAC AAGCTGTGCCAC CCCGAGGAAC TA GTGCTG	CF176 [SEQ ID NO:36] GAACCTCGAGGA TCCTCATTTAGGG CTGGGCAAGGTG CCTTAAGACGCG GTACGACACCTC CAGGAAGCTCTG
T133N, G135T <i>Eco47III</i>	CF177 [SEQ ID NO:37] GTAAGCTTGC GT CGACGCTAGCGG CGCGCCGCCATG GCCGGACCTGCC ACCCAGAGCCCC ATGAAGCTG	T133Nrev [SEQ ID NO:38] GCCCCGGCGCTGG AAAGCGCTGGCG AAGGGCGGCATG GCGGTCTGGTTG GGCTGCAGGGCA G	T133Nfor [SEQ ID NO:39] GGCCCTGCCCC CGAGCCCAACCA GACCGCCATGCC GGCCTTCGCCAG CGCTTTCCAGCG G	CF176 [SEQ ID NO:40] GAACCTCGAGGA TCCTCATTTAGGG CTGGGCAAGGTG CCTTAAGACGCG GTACGACACCTC CAGGAAGCTCTG

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A141N, A143T <i>SapI</i>	CF177 [SEQ ID NO:41] GTAAGCTTGCCT CGACGCTAGCGG CGCGCCGCCATG GCCGGACCTGCC ACCCAGAGCCCC ATGAAGCTG	A141Nrev[SEQ ID NO:42] GCCCGGCGCTGG AAGGTAGAGTTG AAGGGCGGCATG GCACCCCTGGTG GGCTGAAGAGCA GGGGCCAT	A141Nfor[SEQ ID NO:43] GGGAATGGCCCC TGCTCTTCAGCC CACCCAGGGTGC CATGCCGGCCTT CAACTCTACCTT CCAGCGCCGGGC AG	CF176 [SEQ ID NO:44] GAACCTCGAGGA TCCTCATTAGGG CTGGGCAAGGTG CCTTAAGACGCG GTACGACACCTC CAGGAAGCTCTG
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P57V, W58N, P60T <i>HpaI</i>	JCB128 [SEQ ID NO:45] GCTAGCGGCCGCG CCACCATG	JCB136 [SEQ ID NO:46] GCTCAGGGTAGC GTTAACGATGCC CAGAGACTG	JCB137 [SEQ ID NO:47] GGGCATCGTTAA CGCTACCCCTGAG CAGCTG	JCB129 [SEQ ID NO:48] GACTCGAGGATC CTCATTAGGGCT GGG
Q67N, L69T <i>NaeI</i>	JCB134 [SEQ ID NO:49] GCTAGCGGCCGCG CCACCATGGCCG GACCTGCCACCC AG	JCB138 [SEQ ID NO:50] CAAGCAGCCGGC CAGCTGGTGGC GTTGCTGGGCA GCTGCTCAG	JCB139 [SEQ ID NO:51] GCCCCAGCAACG CCACCCAGCTGG CCGGCTGCTTGA G	JCB135 [SEQ ID NO:52] GACTCGAGGATC CTCATTAGGGCT GGGCAAGGTGCC TTAACGACCGG
P60N, S62T <i>SpeI</i>	JCB128 [SEQ ID NO:53] GCTAGCGGCCGCG CCACCATG	JCB130 [SEQ ID NO:54] GGGGCAACTAGT CAGGTTAGCCCA GGG	JCB131 [SEQ ID NO:55] GCTAACCTGACT AGTTGCCCAAGC CAG	JCB129 [SEQ ID NO:56] GACTCGAGGATC CTCATTAGGGCT GGG
S63N, P65T <i>MfeI</i>	JCB128 [SEQ ID NO:57] GCTAGCGGCCGCG CCACCATG	JCB132 [SEQ ID NO:58] GGTGAATTGCT CAGGGGAGCCCA G	JCB133 [SEQ ID NO:59] GCAATTGCACCA GCCAGGCCCTG	JCB129 [SEQ ID NO:60] GACTCGAGGATC CTCATTAGGGCT GGG
E93N, I95T <i>BspEI</i>	JCB134 [SEQ ID NO:61] GCTAGCGGCCGCG CCACCATGGCCG GACCTGCCACCC	JCB140 [SEQ ID NO:62] CCGGACTGGTCC CGTTCAGGGCT GCAGGAGCCCT	JCB141 [SEQ ID NO:63] GAACGGGACCAAG TCCGGAGTTGGG TCCCACCTTGG	JCB135 [SEQ ID NO:64] GACTCGAGGATC CTCATTAGGGCT GGGCAAGGTGCC

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	AG	G		TTAAGACGCGG
<i>Sali</i>	JCB155 [SEQ ID NO:65] GTCGACGCTAGC GGCGCGCCACCA TGGCCGGACTG			

*Nucleotides in bold represent changes imposed in the target sequence and nucleotides in bold *and* italics represent flanking sequences which may add restriction sites to facilitate cloning, Kozac sequences, or stop codons.

Preparation 1a: DNA encoding wild-type human G-CSF

A strand overlapping extension PCR reaction was used to create a wild type human G-CSF construct in order to eliminate the methylation of an ApaI site. Isolated human G-CSF cDNA served as the template for these reactions. The 5' end A primer was used to create a restriction enzyme site prior to the start of the coding region as well as to introduce a Kozac sequence (GGCGCC) 5' of the coding leader sequence to facilitate translation in cell culture.

The A-B product was generated using primers CF177 and CF178 in a PCR reaction. Likewise, the C-D product was produced with primers CF179 and CF176. The products were isolated and combined. The combined mixture was then used as a template with primers CF177 and CF178 to create the full-length wild-type construct. [Nelson, R.M. and Long, G.C. (1989), *Anal. Biochem.* 180:147-151].

The full-length product was ligated into the pCR2.1-Topo vector (Invitrogen, Inc. Cat. No. K4500-40) by way of a topoisomerase TA overhang system to create pCR2.1G-CSF.

The following protocol was used for preparation of the full-length wild-type G-CSF protein as well as each of the G-CSF analogs. Approximately 5 ng of template DNA and 15 pmol of each primer was used in the initial PCR reactions. The reactions were prepared using Platinum PCR Supermix® (GibcoBRL Cat. No. 11306-016). The PCR reactions were

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denatured at 94°C for 5 min and then subject to 25 cycles wherein each cycle consisted of 30 seconds at 94°C followed by 30 seconds at 60°C followed by 30 seconds at 72°C. A final extension was carried out for 7 minutes at 72°C. PCR fragments were isolated from agarose gels and purified using a Qiaquick® gel extraction kit (Qiagen, Cat. No. #28706). DNA was resuspended in sterile water and used for the final PCR reaction to prepare full-length product.

Preparation 1b: DNA encoding G-CSF[A37N,Y39T,P57V,W58N,P60T,Q67N,L69T] was constructed as follows:

DNA encoding G-CSF[A37N,Y39T,Q67N,L69T] was subcloned into pJB02 to create pJB02G-CSF[A37N,Y39T,Q67N,L69T] and pJB02G-CSF[A37N,Y39T,P57V,W58N,P60T] served as the template for strand overlapping expression PCR. JCB155 and JCB136 served as the A and B primers and JCB137 and JCB135 served as the C and D primers. The full-length mutated cDNA was prepared as described previously using JCB155 and JCB134 primers. The resulting full-length DNA encodes a protein with consensus N-linked glycosylation sites in region 1, region 2, and region 9 of the protein (See Figure 1). The full-length cDNA was ligated back into pCR2.1-Topo to create pCR2.1G-CSF[A37N,Y39T,P57V,W58N,P60T,Q67N,L69T].

Preparation 1c: DNA encoding G-CSF[A37N,Y39T,S63N,P64T,E93N,I95T] was constructed as follows:

DNA encoding G-CSF[A37N,Y39T,E93N,I95T] was subcloned into pJB02 to create pJB02G-CSF[A37N,Y39T,E93N,I95T] and pJB02G-CSF[A37N,Y39T,E93N,I95T] served as the template for strand overlapping expression PCR. JCB155 and JCB132 served as the A and B primers and JCB133 and JCB135 served as the C and D primers. The full-length mutated cDNA was prepared as described previously using JCB155 and JCB135 primers. The resulting full-length DNA encodes a protein with consensus N-linked glycosylation sites in region 1, region 7, and

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region 10 of the protein (See Figure 1). The full-length cDNA was ligated back into pCR2.1-Topo to create pCR2.1G-CSF[A37N,Y39T,S63N,P64T,E93N,I95T].

Preparation 1d: DNA encoding G-CSF[C17A] which is G-CSF wherein the amino acid at position 17 is substituted with Ala is constructed as follows:

The wild-type construct in the pCR2.1-Topo vector (pCR2.1G-CSF) serves as the PCR template for the C17A mutagenesis. Strand overlapping extension PCR is performed as described previously. CF177 and C17Arev serve as the A-B primers and C17Afor and CF176 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the CF177 and CF176 primers. The B and C primers are used to mutate the DNA such that a *SacI* restriction site is created and the protein expressed from the full-length sequence contains an Alanine instead of a Cysteine at position 17. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[C17A] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe*/Xho sites of mammalian expression vector pJB02 to create pJB02G-CSF[C17A].

Preparation 1e: DNA encoding G-CSF[A37N,Y39T] is constructed as follows:

Strand overlapping extension PCR is performed using pCR2.1G-CSF[C17A] as the template. Primers CF177 and A37Nrev serve as the A-B primers and CF176 and A37Nfor serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the CF177 and CF176 primers. The B and C primers contain mismatched sequences such that a *SpeI* site is created in the DNA and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 1 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[A37N,Y39T] wherein the sequence is

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confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe/Xho* sites of mammalian expression vector pJB02 to create pJB02G-CSF[A37N,Y39T].

Preparation 1f: DNA encoding G-CSF[P57V,W58N,P60T] is constructed as follows:

Strand overlapping extension PCR is performed using pJB02G-CSF[C17A] as the template. Primers JCB128 and JCB136 serve as the A-B primers and JCB137 and JCB129 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the JCB128 and JCB129 primers. The B and C primers contain mismatched sequences such that a *HpaI* site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 2 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[P57V,W58N,P60T] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe/Xho* sites of mammalian expression vector pJB02 to create pJB02G-CSF[P57V,W58N,P60T].

Preparation 1g: DNA encoding G-CSF[P60N,S62T] is constructed as follows:

Strand overlapping extension PCR is performed using pJB02G-CSF[C17A] as the template. Primers JCB128 and JCB130 serve as the A-B primers and JCB131 and JCB129 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the JCB128 and JCB129 primers. The B and C primers contain mismatched sequences such that a *SpeI* site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 4 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[P60N,S62T] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the

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Nhe/Xho sites of mammalian expression vector pJB02 to create pJB02G-CSF[P60N,S62T].

Preparation 1h: DNA encoding G-CSF[S63N,P65T] is constructed as follows:

Strand overlapping extension PCR is performed using pJB02G-CSF[C17A] as the template. Primers JCB128 and JCB132 serve as the A-B primers and JCB133 and JCB129 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the JCB128 and JCB129 primers. The B and C primers contain mismatched sequences such that a *MfeI* site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 7 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[S63N,P65T] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe/Xho* sites of mammalian expression vector pJB02 to create pJB02G-CSF[S63N,P65T].

Preparation 1i: DNA encoding G-CSF[Q67N,L69T] is constructed as follows:

Strand overlapping extension PCR is performed using pJB02G-CSF[C17A] as the template. Primers JCB134 and JCB138 serve as the A-B primers and JCB139 and JCB135 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the JCB128 and JCB129 primers. The B and C primers contain mismatched sequences such that a *NaeI* site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 9 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[Q67N,L69T] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe/Xho* sites of mammalian expression vector pJB02 to create pJB02G-CSF[Q67N,L69T].

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Preparation 1j: DNA encoding G-CSF[E93N,I95T] is constructed as follows:

Strand overlapping extension PCR is performed using pJB02G-CSF[C17A] as the template. Primers JCB134 and JCB140 serve as the A-B primers and JCB141 and JCB135 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the JCB128 and JCB129 primers. The B and C primers contain mismatched sequences such that a *BspEI* site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 10 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[E93N,I95T] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe/Xho* sites of mammalian expression vector pJB02 to create pJB02G-CSF[E93T,I95T].

Preparation 1k: DNA encoding G-CSF[T133N,G135T] is constructed as follows:

Strand overlapping extension PCR is performed using pCR2.1G-CSF[C17A] as the template. Primers CF177 and T133Nrev serve as the A-B primers and T133Nfor and CF176 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the CF177 and CF176 primers. The B and C primers contain mismatched sequences such that an *Eco47III* site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 13 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[T133N,G135T] wherein the sequence is confirmed.

Preparation 1l: DNA encoding G-CSF[A141N,A143T] is constructed as follows:

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Strand overlapping extension PCR is performed using pCR2.1G-CSF[C17A] as the template. Primers CF177 and A141Nrev serve as the A-B primers and A141Nfor and CF176 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the CF177 and CF176 primers. The B and C primers contain mismatched sequences such that an *SapI* site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 14 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[A141N,A143T] wherein the sequence is confirmed.

Preparation 1m: DNA encoding G-CSF[A37N,Y39T,T133N,G135T] is constructed as follows:

A 210 bp insert containing G-CSF[A37N,Y39T] is isolated from pCR2.1G-CSF[A37N,Y39T] using *EcoNI*. This fragment is ligated into pCR2.1G-CSF[T133N,G135T] which is prepared by cleavage with *EcoNI* and subsequent isolation of the vector (4359 bp) from a 210 bp fragment containing wild-type G-CSF sequences. This ligation creates pCR2.1G-CSF[A37N,Y39T,T133N,G135T]. Analog encoding DNA is then subcloned into pJB02 using *NheI/XhoI* to create pJB02G-CSF[A37N,Y39T,T133N,G135T].

Preparation 1n: DNA encoding G-CSF[A37N,Y39T,A141N,A143T] is constructed as follows:

A 210 bp insert containing G-CSF[A37N,Y39T] is isolated from pCR2.1G-CSF[A37N,Y39T] using *EcoNI*. This fragment is ligated into pCR2.1G-CSF[A141N,A143T] which is prepared by cleavage with *EcoNI* and subsequent isolation of the vector (4359 bp) from a 210 bp fragment containing wild-type G-CSF sequences. This ligation creates pCR2.1G-CSF[A37N,Y39T,A141N,A143T]. Analog encoding DNA is then subcloned into pJB02 (Figure 3) using *NheI/XhoI* to create pJB02G-CSF[A37N,Y39T,A141N,A143T].

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Preparation 1o: DNA encoding G-CSF[A37N,Y39T,P57V,W58N,P60T] is constructed as follows:

DNA encoding G-CSF[A37N,Y39T] is subcloned into pJB02 to create pJB02G-CSF[A37N,Y39T] and pJB02G-CSF[A37N,Y39T] serves as the template for strand overlapping expression PCR. JCB128 and JCB136 serve as the A and B primers and JCB137 and JCB129 serve as the C and D primers. The full-length mutated cDNA is prepared as described previously using JCB128 and JCB129 primers. The resulting full-length DNA encodes a protein with consensus N-linked glycosylation sites in region 1 and region 2 of the protein. The full-length cDNA is ligated back into pCR2.1-Topo to create pCR2.1G-CSF[A37N,Y39T,P57V,W58N,P60T].

Preparation 1p: DNA encoding G-CSF[A37N,Y39T,Q67N,L69T] is constructed as follows:

DNA encoding G-CSF[A37N,Y39T] is subcloned into pJB02 to create pJB02G-CSF[A37N,Y39T] and pJB02G-CSF[A37N,Y39T] serves as the template for strand overlapping expression PCR. JCB134 and JCB138 serve as the A and B primers and JCB139 and JCB135 serve as the C and D primers. The full-length mutated cDNA is prepared as described previously using JCB128 and JCB129 primers. The resulting full-length DNA encodes a protein with consensus N-linked glycosylation sites in region 1 and region 9 of the protein. The full-length cDNA is ligated back into pCR2.1-Topo to create pCR2.1G-CSF[A37N,Y39T,Q67N,L69T].

Preparation 1q: DNA encoding G-CSF[A37N,Y39T,E93N,I95T] is constructed as follows:

DNA encoding G-CSF[A37N,Y39T] is subcloned into pJB02 to create pJB02G-CSF[A37N,Y39T] and pJB02G-CSF[A37N,Y39T] serves as the template for strand overlapping expression PCR. JCB134 and JCB140 serve as the A and B primers and JCB141 and JCB135 serve as the C and D primers. The full-

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length mutated cDNA is prepared as described previously using JCB128 and JCB129 primers. The resulting full-length DNA encodes a protein with consensus N-linked glycosylation sites in region 1 and region 10 of the protein. The full-length cDNA is ligated back into pCR2.1-Topo to create pCR2.1G-CSF[A37N,Y39T,E93N,I95T].

Example 2: Expression of heterologous fusion proteins:

2a: Expression in 293/EBNA cells:

Each full-length DNA encoding a G-CSF analog was subcloned into the *NheI/XhoI* sites of mammalian expression vector pJB02 (Figure 3). This vector contains both the Ori P and Epstein Barr virus nuclear antigen (EBNA) components which are necessary for sustained, transient expression in 293 EBNA cells. This expression plasmid contains a puromycin resistance gene expressed from the CMV promoter as well as an ampicillin resistance gene. The gene of interest is also expressed from the CMV promoter.

The transfection mixture was prepared by mixing 73 µl of the liposome transfection agent Fugene 6® (Roche Molecular Biochemicals, Cat. No. 1815-075) with 820 µl Opti-Mem® (GibcoBRL Cat. No. 31985-062). G-CSF pJB02 DNA (12µg), prepared using a Qiagen plasmid maxiprep kit (Qiagen, Cat. No. 12163), was then added to the mixture. The mixture was incubated at room temperature for 15 minutes.

Cells were plated on 10 cm² plates in DMEM/F12 3:1 (GibcoBRL Cat. No. 93-0152DK) supplemented with 5% fetal bovine serum, 20mM HEPES, 2 mM L-glutamine, and 50 µg/mL Geneticin such that the plates were 60% to 80% confluent by the time of the transfection. Immediately before the transfection mixture was added to the plates, fresh media was added. The mixture was then added dropwise to cells with intermittent swirling. Plates were then incubated at 37°C in a 5% CO₂ atmosphere for 24 hours at which point the media was changed to Hybritech medium without serum. The

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media containing a secreted form of a glycosylated G-CSF analog was then isolated 48 hours later.

2b: Expression in CHO cells:

The expression vector for expression in CHO-K1 cells pEE14.1 is illustrated in Figure 4. This vector includes the glutamine synthetase gene which enables selection using methionine sulfoximine. This gene includes two poly A signals at the 3' end. G-CSF analogs are expressed from the CMV promoter which includes 5' untranslated sequences from the hCMV-MIE gene to enhance mRNA levels and translatability. The SV40 poly A signal is cloned 3' of the G-CSF analog DNA. The SV40 late promoter drives expression of GS minigene. This expression vector encoding the gene of interest was prepared for transfection using a QIAGEN Maxi Prep Kit (QIAGEN, Cat. No. 12362). The final DNA pellet (50-100 µg) was resuspended in 100 µl of basal formulation medium (GibcoBRL CD-CHO Medium without L-Glutamine, without thymidine, without hypoxanthine). Before each transfection, CHO-K1 cells were counted and checked for viability. A volume equal to 1×10^7 cells was centrifuged and the cell pellet rinsed with basal formulation medium. The cells were centrifuged a second time and the final pellet resuspended in basal formulation medium (700 µl final volume).

The resuspended DNA and cells were then mixed together in a standard electroporation cuvette (Gene Pulsar Cuvette) used to support mammalian transfections, and placed on ice for five minutes. The cell/DNA mix was then electroporated in a BioRad Gene Pulsar device set at 300V/975 µF and the cuvette placed back on ice for five minutes. The cell/DNA mixed was then diluted into 20 ml of cell growth medium in a non-tissue culture treated T75 flask and incubated at 37°C / 5% CO₂ for 48-72 hours.

The cells were counted, checked for viability, and plated at various cell densities in selective medium in 96

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well tissue culture plates and incubated at 37°C in a 5% CO₂ atmosphere. Selective medium is basal medium with 1X HT Supplement (GibcoBRL 100X HT Stock), 100 µg/mL Dextran Sulfate (Sigma 100 mg/ml stock), 1X GS Supplements (JRH BioSciences 50X Stock) and 25 µM MSX (Methionine Sulphoximine). The plates were monitored for colony formation and screened for glycosylated G-CSF analog production.

Example 3: Purification of Heterologous Fusion Proteins

HA Fusions

The cell culture harvest was dialyzed against 20 mM Tris pH 7.4. An anion exchange column (1 ml Pharmacia HiTrap Q) was equilibrated with 20 mM Tris pH 7.4 and the dialyzed material loaded at 2 ml/min. The protein was eluted from the column using a linear gradient from 0 to 500 mM NaCl in 80 min at 1 ml/min and elution was monitored by UV absorbance at 280 nm. SDS-PAGE analysis was used to identify and pool fractions of interest. This pool was dialyzed against 25 mM sodium acetate (NaOAc) pH 5.0

A cation exchange column (1 ml Pharmacia HiTrap S column) was equilibrated with 25 mM NaOAc pH 5.0 and the dialysate was loaded at 1 ml/min. The protein was eluted from the column using a linear gradient from 0 to 500 mM NaCl in 30 min. The fractions were immediately neutralized with 1 M Tris pH 8 to a final pH of 7. SDS-PAGE gels were used to identify and pool fractions of interest.

Fc Fusions

The cell culture harvest was dialyzed against 20 mM sodium phosphate pH 7.0. An affinity column (1 ml Pharmacia HiTrap Protein A or rProtein A) was equilibrated with 20 mM sodium phosphate pH 7.0 and the dialysate was loaded at 2 ml/min. 1 ml/min of 100 mM citric acid pH 3 was used to

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elute the protein. Fractions were immediately neutralized with 1M Tris pH 8 to pH 7 and peak fractions (determined by in-line OD280 monitoring) were further diluted with 20 mM sodium phosphate pH 7.0. SDS-PAGE analysis was used to identify and pool fractions of interest.

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WE CLAIM:

1. A heterologous fusion protein comprising a hyperglycosylated G-CSF analog fused to a polypeptide selected from the group consisting of
 - d) human albumin;
 - e) human albumin analogs; and
 - f) fragments of human albumin.
2. The heterologous fusion protein of claim 1, wherein the hyperglycosylated G-CSF analog is fused to the polypeptide via a peptide linker.
3. The heterologous fusion protein of the Claim 2 wherein the peptide linker is selected from the group consisting of:
 - c) a glycine rich peptide;
 - d) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]_n where n is 1, 2, 3, 4, or 5; and
 - e) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]₃.
4. The heterologous fusion protein of Claims 1, 2, or 3 wherein the hyperglycosylated G-CSF analog comprises the amino acid sequence of the formula I: [SEQ ID NO: 1]

1	5	10	15
Thr Pro Leu Gly Pro Ala Ser Ser	Leu Pro Gln Ser Phe Leu	Leu Lys	
20	25	30	
Xaa Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly	Ala Ala Leu Gln		
35	40	45	
Glu Lys Leu Cys Xaa Xaa Lys Leu Cys His Pro	Glu Glu Leu Val		
50	55	60	
Leu Leu Gly His Ser Leu Gly Ile Xaa Xaa Xaa Xaa Xaa	Xaa Xaa Xaa Xaa		
65	70	75	80
Xaa Xaa Xaa Xaa Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser			
85	90	95	
Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Xaa Xaa Ser			
100	105	110	
Xaa Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp			
115	120	125	
Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro			

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130 135 140
Ala Leu Gln Pro Xaa Xaa Xaa Ala Met Pro Ala Phe Xaa Xaa Xaa Phe
145 150 155 160
Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Aln Ser Phe
165 170
Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro (I)

wherein:

Xaa at position 17 is Cys, Ala, Leu, Ser, or Glu;
Xaa at position 37 is Ala or Asn;
Xaa at position 38 is Thr, or any other amino acid except Pro;
Xaa at position 39 is Tyr, Thr, or Ser;
Xaa at position 57 is Pro or Val;
Xaa at position 58 is Trp or Asn;
Xaa at position 59 is Ala or any other amino acid except Pro;
Xaa at position 60 is Pro, Thr, Asn, or Ser,
Xaa at position 61 is Leu, or any other amino acid except Pro;
Xaa at position 62 is Ser or Thr;
Xaa at position 63 is Ser or Asn;
Xaa at position 64 is Cys or any other amino acid except Pro;
Xaa at position 65 is Pro, Ser, or Thr;
Xaa at position 66 is Ser or Thr;
Xaa at position 67 is Gln or Asn;
Xaa at position 68 is Ala or any other amino acid except Pro;
Xaa at position 69 is Leu, Thr, or Ser
Xaa at position 93 is Glu or Asn
Xaa at position 94 is Gly or any other amino acid except Pro;
Xaa at position 95 is Ile, Asn, Ser, or Thr;
Xaa at position 97 is Pro, Ser, Thr, or Asn;
Xaa at position 133 is Thr or Asn;
Xaa at position 134 is Gln or any other amino acid except Pro;
Xaa at position 135 is Gly, Ser, or Thr
Xaa at position 141 is Ala or Asn;
Xaa at position 142 is Ser or any other amino acid except Pro; and
Xaa at position 143 is Ala, Ser, or Thr;

and wherein:

Xaa at positions 37, 38, and 39 constitute region 1;
Xaa at positions 58, 59, and 60 constitute region 2;
Xaa at positions 59, 60, and 61 constitute region 3;

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Xaa at positions 60, 61, and 62 constitute region 4;
Xaa at positions 61, 62, and 63 constitute region 5;
Xaa at positions 62, 63, and 64 constitute region 6;
Xaa at positions 63, 64, and 65 constitute region 7;
Xaa at positions 64, 65, and 66 constitute region 8;
Xaa at positions 67, 68, and 69 constitute region 9;
Xaa at positions 93, 94, and 95 constitute region 10;
Xaa at positions 94, 95, and Ser at position 96
constitute region 11;
Xaa at positions 95, and 97, and Ser at position 96
constitute region 12;
Xaa at positions 133, 134, and 135 constitute
region 13;
Xaa at positions 141, 142, and 143 constitute
region 14;

and provided that at least one of regions 1 through 14
comprises the sequence Asn Xaa1 Xaa2 wherein Xaa1 is any
amino acid except Pro and Xaa2 is Ser or Thr.

5. The heterologous fusion protein of Claim 4 wherein any
two regions of regions 1 through 14 comprise the sequence
Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and
Xaa2 is Ser or Thr.

6. The heterologous fusion protein of Claim 4 wherein any
three regions of regions 1 through 14 comprise the sequence
Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and
Xaa2 is Ser or Thr.

7. The heterologous fusion protein of Claim 4 wherein any
four regions of regions 1 through 14 comprise the sequence
Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and
Xaa2 is Ser or Thr.

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8. The heterologous fusion protein of Claim 4 wherein the hyperglycosylated G-CSF analog is selected from the group consisting of:

- a) G-CSF [A37N, Y39T]
- b) G-CSF [P57V, W58N, P60T]
- c) G-CSF [P60N, S62T]
- d) G-CSF [S63N, P65T]
- e) G-CSF [Q67N, L69T]
- f) G-CSF [E93N, I95T]
- g) G-CSF [T133N, G135T]
- h) G-CSF [A141N, A143T]
- i) G-CSF [A37N, Y39T, P57V, W58N, P60T]
- j) G-CSF [A37N, Y39T, P60N, S62T]
- k) G-CSF [A37N, Y39T, S63N, P65T]
- l) G-CSF [A37N, Y39T, Q67N, L69T]
- m) G-CSF [A37N, Y39T, E93N, I95T]
- n) G-CSF [A37N, Y39T, T133N, G135T]
- o) G-CSF [A37N, Y39T, A141N, A143T]
 - p) G-CSF [A37N, Y39T, P57V, W58N, P60T, S63N, P65T]
- q) G-CSF [A37N, Y39T, P57V, W58N, P60T, Q67N, L69T]
- r) G-CSF [A37N, Y39T, S63N, P65T, E93N, I95T].

9. The heterologous fusion protein of claim 8, wherein the hyperglycosylated G-CSF analog is G-CSF [A37N, Y39T, P57V, W58N, P60T, Q67N, L69T].

10. The heterologous fusion protein of claim 8, wherein the hyperglycosylated G-CSF analog is G-CSF [A37N, Y39T, S63N, P65T, E93N, I95T].

11. A heterologous fusion protein which is the product of the expression in a host cell of an exogenous DNA sequence which comprises a DNA sequence encoding a heterologous fusion protein of any one of Claims 1 through 11.

12. An isolated nucleic acid sequence, comprising a

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polynucleotide encoding a heterologous fusion protein of any one of Claims 1 through 11.

13. An isolated nucleic acid sequence, comprising a polynucleotide which comprises a DNA sequence selected from the group consisting of:

a) SEQ ID NO:2

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG CTC TTC GAC ACA CGG TGG ATG TTC GAC ACG GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC AGC GAC GAG CCT GTG ACA GAC CCG TAG GGG ACC CGA GGG GAC TCG TCG ACG

CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC GGG TCG GTC CGG GAC GTC GAC CGT CCG ACG AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC CGG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTC CAG CTG GAC GTC GCC GAC

GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC AAC CAG ACC GCC ATG CCG GCC TTC GCC TCT GCT TTC CGG GAC GTC GGG TTG GTC TGG CGG TAC GGC CGG AAG CGG AGA CGA AAG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC GAC CTC CAC AGC ATG CGC CAG AAT TCC GTG GAA CGG GTC CGG

b) SEQ ID NO:3

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG CTC TTC GAC ACA CGG TGG ATG TTC GAC ACG GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC AGC GAC GAG CCT GTG ACA GAC CCG TAG GGG ACC CGA GGG GAC TCG TCG ACG

CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC

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GGG TCG GTC CGG GAC GTC GAC CGT CCG ACG AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC CAG GAC GTC CGG GAC CTT CCC TAG AGGCCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TTG AAC CTG TGT GAC GTC CGT CAG CGG CTGTTT GCC ACC ACC ATC TTG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGAGCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC AAC TCT ACC TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG TTG AGA TGG AAGCAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGC GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAGCTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGC CAG AAT TCC GTG GAA CGG GTC GGG

c) SEQ ID NO:4

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTCGCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA CGC CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTCGAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC AGC GTG GGG CTC CTC GAC CACCTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC AGC TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TCG AGCCCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CCT CGG AGC AAC TCG GTT GAG GTA TCGGGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC CAG GAC GTC CGG GAC CTT CCC TAG AGGCCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC CGC GAC CTG CAG CGG CTGTTT GCC ACC ACC ATC TTG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGAGCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAGCAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGC GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAGCTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGC CAG AAT TCC GTG GAA CGG GTC GGG

d) SEQ ID NO:5

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

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GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CGG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
GTC TTC GAC ACA CGG TGG ATG TTC GAC ACG GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT AAC ACT AGC AGG TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA TTG GAC TCC TCG ACC

CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TGC CGG GAC GTC GAC CGT CGG ACG AAC TCG TTG GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA CGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CGG GCC TTC GCT CCT GCT TTC
CGG GAC GTC CGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG

CAG CGC CGG GCA GGA GGG GTC CTG TTG GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CGC
GAC CTC CAC AGC ATG CGG CAG AAT TCC GTG GAA CGG GTC GGG

e) SEQ ID NO: 5
ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TGG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CGG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC ACG GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AAT TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TTA ACG

ACC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
TGG TCG GTC CGG GAC GTC GAC CGT CGG ACG AAC TCG TTG GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA CGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CGG GCC TTC GCT CCT GCT TTC
CGG GAC GTC CGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG

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CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC GCG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGG CAG AAT TCC GTG GAA CGG GTC GGG

f) SEQ ID NO:7

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CCT CGC GAG GTC

GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC GTT AAC GCT ACC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CGG TAG CAA TTG CGA TGG GAC TCG TCG ACG

CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CCT CGG ACG AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CCT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA CGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CCT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CGG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAC CGG AGA CGA AAG

CAG CCC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC GCG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGG CAG AAT TCC GTG GAA CGG GTC GGG

g) SEQ ID NO:8

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CCT CGC GAG GTC

GAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TCG ACG

CCC AGC AAC GCC ACC CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG TTG CGG TGG GTC GAC CGT CCT CGG ACG AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC

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CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTGTTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC TAC CTT GAC CCT TAC CGG GGAGCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAGCAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAGCTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC CGG

h) SEQ ID NO:9

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTCGCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTCGAG AAG CTG TGT GCC ACC TAC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA CGG TGG ATG TTC GAC AGC GTG GGG CTC CTC GAC CACCTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC AGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TCG AGCCCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CCT CGG AGC AAC TCG GTT GAG GTA TCGGGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG AAC GGG ACC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC TTG CCC TGG AGGCCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTGTTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC TAC CTT GAC CCT TAC CGG GGAGCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAGCAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAGCTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC CGG

i) SEQ ID NO:10

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTCGCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

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GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TCG AGC

CCC AGC CGG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT ACC
GGG TCG GTC CGG GAC GTC CGT CGG AGC AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTC GAC CCT TAC CGG GGA

GCC CTG CAG CCC AAC CAG ACC GCC ATC CGG GCC CCC TTG GCC TCT GCT
CGG CTC AAC CCA GGG TGG TGG CGG TAG ACC GCA CAA CGG AGG GTA GAC GTC CGG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG CAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG CGG CAG AAT TCC GTG GAA CGG GTC GGG

j) SEQ ID NO:11
ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GTC TTC

GCC CTG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC GTC CGG CTA CGG CGT CGG GAG GTC

GAG AAG CTG TGT AAC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC AGC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GCC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TCG AGC

GCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC CGT CGG AGC AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CGG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTC GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CGG GCC TTC AAC TCT ACC TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AGG TTG AGA TGG AAG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

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CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC GGG

k) SEQ ID NO:12

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGC GAC ACC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC GTT AAC GCT ACC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCG TAG CAA TTG CGA TGG GAC TCG TCG AGC

CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC GAC CGT CCG ACG AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTC GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GGC CCT
AAA CGG TGG TAG ACC GTC GTC TAC CCT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CCG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG

CAG CCC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC GCG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC GGG

l) SEQ ID NO:13

ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGC GAC ACC GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCG TAG GGG ACC CGA GGG GAC TCG TCG ACC

CCC AGC AAC GCC ACC CAG CTG GCA GGG TGC TTG AGC CAA CTC CAT AGC
GGG TCG TTG CGG TGG GTC GAC CGT CCG ACG AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG

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CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CCG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC GCG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC CGG

m) SEQ ID NO:14
ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA CGG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC ACG GTG GGG CTC CTC GAC CAC

CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCG TAG GGG ACC CGA GGG GAC TCG TCG ACG

CCC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG GTC CGG GAC GTC CAC CGT CCT CGG ACG AAC TCG GTT GAG GTA TCG

GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GGC CTG GAA GGG ATC TCC
CCG GAA AAG GAG ATG GTC CCC GAG GAC GTC CCG GAC CTT CCC TAG AGG

AAC GGT ACC GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
TTG CCA TGG CCA CGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG

TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT CTT GAC CCT TAC CGG GGA

GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CCG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG

CAG CGC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC GCG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG

CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC CGG

n) SEQ ID NO:15
ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CGG GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC

GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA CGG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC

GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC ACG GTG GGG CTC CTC GAC CAC

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CTG CTC GGA CAC TCT CTG GGC ATC GTT AAC GCT ACC CTG AGC AGC TGC
GAC GAG CCT GTG ACA GAC CCG TAG CAA TTG CGA TGG GAC TCG TCG AGC
CCC AGC AAC GCC ACC CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
GGG TCG TTG CGG TGG GTC GAC CGT CCC AGC AAC TCG GTT GAG GTA TCG
GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG ATC TCC
CCC GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC CTT CCC TAG AGG
CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CGG CGG CTG
TTT GCC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG ACC GTC GTC TAC CTT GAC CCT TAC CGG GGA
GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG
CAG CCC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG
CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC GGG
o) SEQ ID NO:16
ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG AGC TTC CTG CTC AAG
TGG GGG GAC CCC GGA CGG TCG AGG GAC GGG GTC TCG AAG GAC GAG TTC
GCC TTA GAG CAA GTG AGG AAG ATC CAG GGC GAT GGC GCA GCG CTC CAG
CGG AAT CTC GTT CAC TCC TTC TAG GTC CCG CTA CGG CGT CGC GAG GTC
GAG AAG CTG TGT AAC ACC ACC AAG CTG TGC CAC CCC GAG GAG CTG GTG
CTC TTC GAC ACA TTG TGG TGG TTC GAC AGC GTG GGG CTC CTC GAC CAC
CTG CTC GGA CAC TCT CTG GGC ATC CCC TGG GCT CCC CTG AGC AAT TGC
GAC GAG CCT GTG ACA GAC CGG TAG GGG ACC CGA GGG GAC TCG TTA AGC
ACC AGC CAG GCC CTG CAG CTG GCA GGC TGC TTG AGC CAA CTC CAT AGC
TGG TCG GTC CGG GAC GTC GAC CGT CCC AGC AAC TCG GTT GAG GTA TCG
GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG AAC GGG ACC TCC
CGG GAA AAG GAG ATG GTC CCC GAG GAC GTC CGG GAC TTG CCC TGG AGG
CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC
GGG CTC AAC CCA GGG TGG AAC CTG TGT GAC GTC GAC CTG CAG CGG CTG
TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT
AAA CGG TGG TGG TAG ACC GTC GTC TAC CTT GAC CCT TAC CGG GGA
GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC
CGG GAC GTC GGG TGG GTC CCA CGG TAC GGC CGG AAG CGG AGA CGA AAG
CAG CCC CGG GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC
GTC CGG GCC CGT CCT CCC CAG GAC CAA CGG AGG GTA GAC GTC TCG AAG
CTG GAG GTG TCG TAC CGC GTC TTA AGG CAC CTT GCC CAG CCC
GAC CTC CAC AGC ATG GCG CAG AAT TCC GTG GAA CGG GTC GGG,

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fused to a DNA encoding a protein selected from the group consisting of:

- a)human albumin,
- b)human albumin analog; and
- c)fragments of human albumin.

14. The heterologous fusion protein of any one of Claims 1 through 11 wherein the polypeptide is human albumin.

15. The heterologous fusion protein of Claim 12, wherein the second polypeptide has the sequence of SEQ ID NO: 35.

16. The heterologous fusion protein of any one of Claims 1 through 11 wherein the second polypeptide is an N-terminal fragment of albumin.

17. A method for increasing neutrophil levels in a mammal comprising the administration of a therapeutically effective amount of the heterologous fusion protein of any one of Claims 1 through 11, 14 and 15.

18. The use of the heterologous fusion protein as claimed in any one of Claims 1 through 11, 14 and 15 for the manufacture of a medicament for the treatment of patients with insufficient circulating neutrophil levels.

19. Use of a heterologous fusion protein of any one of Claims 1 through 11, 14, and 15 as a medicament.

20. Use of a heterologous fusion protein of any one of Claims 1 through 11, 14, and 15 for the treatment of patients with insufficient circulating neutrophil levels.

21. A pharmaceutical formulation adapted for the treatment of patients with insufficient neutrophil levels comprising a

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heterologous fusion protein of any one of Claims 1 through 11, 14, and 15.

22. A heterologous fusion protein comprising a hyperglycosylated G-CSF analog fused to a polypeptide selected from the group consisting of
a) the Fc portion of an immunoglobulin;
b) an analog of the Fc portion of an immunoglobulin;
and
c) fragments of the Fc portion of an immunoglobulin.

23. The heterologous fusion protein of Claim 22, wherein the hyperglycosylated G-CSF analog is fused to the polypeptide via a peptide linker.

24. The heterologous fusion protein of the Claim 23 wherein the peptide linker is selected from the group consisting of:
a) a glycine rich peptide;
b) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]_n where n is 1, 2, 3, 4, or 5; and
c) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]₃.

23. The heterologous fusion protein of Claims 22, 23 or 24, wherein the hyperglycosylated G-CSF analog comprises the amino acid sequence of the formula I: [SEQ ID NO: 1]

1	5	10	15
Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Lys			
20	25	30	
Xaa Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln			
35	40	45	
Glu Lys Leu Cys Xaa Xaa Xaa Lys Leu Cys His Pro Glu Glu Leu Val			
50	55	60	
Leu Leu Gly His Ser Leu Gly Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa			
65	70	75	80
Xaa Xaa Xaa Xaa Xaa Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser			
85	90	95	
Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Xaa Xaa Xaa Ser			
100	105	110	
Xaa Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp			
115	120	125	
Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro			
130	135	140	
Ala Leu Gln Pro Xaa Xaa Xaa Ala Met Pro Ala Phe Xaa Xaa Xaa Phe			
145	150	155	160

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Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Ala Ser Phe
165 170
Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro (I)

wherein:

Xaa at position 17 is Cys, Ala, Leu, Ser, or Glu;
Xaa at position 37 is Ala or Asn;
Xaa at position 38 is Thr, or any other amino acid except Pro;
Xaa at position 39 is Tyr, Thr, or Ser;
Xaa at position 57 is Pro or Val;
Xaa at position 58 is Trp or Asn;
Xaa at position 59 is Ala or any other amino acid except Pro;
Xaa at position 60 is Pro, Thr, Asn, or Ser,
Xaa at position 61 is Leu, or any other amino acid except Pro;
Xaa at position 62 is Ser or Thr;
Xaa at position 63 is Ser or Asn;
Xaa at position 64 is Cys or any other amino acid except Pro;
Xaa at position 65 is Pro, Ser, or Thr;
Xaa at position 66 is Ser or Thr;
Xaa at position 67 is Gln or Asn;
Xaa at position 68 is Ala or any other amino acid except Pro;
Xaa at position 69 is Leu, Thr, or Ser
Xaa at position 93 is Glu or Asn
Xaa at position 94 is Gly or any other amino acid except Pro;
Xaa at position 95 is Ile, Asn, Ser, or Thr;
Xaa at position 97 is Pro, Ser, Thr, or Asn;
Xaa at position 133 is Thr or Asn;
Xaa at position 134 is Gln or any other amino acid except Pro;
Xaa at position 135 is Gly, Ser, or Thr
Xaa at position 141 is Ala or Asn;
Xaa at position 142 is Ser or any other amino acid except Pro; and
Xaa at position 143 is Ala, Ser, or Thr;

and wherein:

Xaa at positions 37, 38, and 39 constitute region 1;
Xaa at positions 58, 59, and 60 constitute region 2;
Xaa at positions 59, 60, and 61 constitute region 3;
Xaa at positions 60, 61, and 62 constitute region 4;

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Xaa at positions 61, 62, and 63 constitute region 5;
Xaa at positions 62, 63, and 64 constitute region 6;
Xaa at positions 63, 64, and 65 constitute region 7;
Xaa at positions 64, 65, and 66 constitute region 8;
Xaa at positions 67, 68, and 69 constitute region 9;
Xaa at positions 93, 94, and 95 constitute region 10;
Xaa at positions 94, 95, and Ser at position 96
constitute region 11;
Xaa at positions 95, and 97, and Ser at position 96
constitute region 12;
Xaa at positions 133, 134, and 135 constitute
region 13;
Xaa at positions 141, 142, and 143 constitute
region 14;

and provided that at least one of regions 1 through 14
comprises the sequence Asn Xaa1 Xaa2 wherein Xaa1 is any
amino acid except Pro and Xaa2 is Ser or Thr.

26. The heterologous fusion protein of Claim 25 wherein any
two regions of regions 1 through 14 comprise the sequence
Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and
Xaa2 is Ser or Thr.

27. The heterologous fusion protein of Claim 25 wherein any
three regions of regions 1 through 14 comprise the sequence
Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and
Xaa2 is Ser or Thr.

28. The heterologous fusion protein of Claim 25 wherein any
four regions of regions 1 through 14 comprise the sequence
Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and
Xaa2 is Ser or Thr.

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29. The heterologous fusion protein of Claim 25 wherein the hyperglycosylated G-CSF analog is selected from the group consisting of:

- a) G-CSF[A37N,Y39T]
- b) G-CSF[P57V,W58N,P60T]
- c) G-CSF[P60N,S62T]
- d) G-CSF[S63N,P65T]
- e) G-CSF[Q67N,L69T]
- f) G-CSF[E93N,I95T]
- g) G-CSF[T133N,G135T]
- h) G-CSF[A141N,A143T]
- i) G-CSF[A37N,Y39T,P57V,W58N,P60T]
- j) G-CSF[A37N,Y39T,P60N,S62T]
- k) G-CSF[A37N,Y39T,S63N,P65T]
- l) G-CSF[A37N,Y39T,Q67N,L69T]
- m) G-CSF[A37N,Y39T,E93N,I95T]
- n) G-CSF[A37N,Y39T,T133N,G135T]
- o) G-CSF[A37N,Y39T,A141N,A143T]
- p) G-CSF[A37N,Y39T,P57V,W58N,P60T,S63N,P65T]
- q) G-CSF[A37N,Y39T,P57V,W58N,P60T,Q67N,L69T]
- r) G-CSF[A37N,Y39T,S63N,P65T,E93N,I95T].

30. The heterologous fusion protein of Claim 29 wherein the hyperglycosylated G-CSF analog is G-CSF[A37N,Y39T,P57V,W58N,P60T,Q67N,L69T].

31. The heterologous fusion protein of Claim 29 wherein the hyperglycosylated G-CSF analog is G-CSF[A37N,Y39T,S63N,P65T,E93N,I95T].

32. The heterologous fusion protein of any one of Claims 22 through 31 wherein the second polypeptide is the Fc portion of an Ig selected from the group consisting of: IgG1, IgG2, IgG3, IgG4, IgE, IgA, IgD, or IgM.

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33. The heterologous fusion protein of any one of Claims 22 through 32 wherein the second polypeptide is the Fc portion of an Ig selected from the group consisting of: IgG1, IgG2, IgG3, and IgG4.

34. The heterologous fusion protein of Claim 33 wherein the second polypeptide is the Fc portion of an IgG1 immunoglobulin.

35. The heterologous fusion protein of Claim 33 wherein the second polypeptide is the Fc portion of an IgG4 immunoglobulin.

36. The heterologous fusion protein of Claims 22 through 35 wherein the IgG is human.

37. The heterologous fusion protein of any one of Claims 22 through 36 wherein the Fc portion comprises the hinge, CH2, and CH3 domains.

38. The heterologous fusion protein of Claim 34 wherein the polypeptide has the sequence of SEQ ID NO: 33.

39. The heterologous fusion protein of Claim 34, wherein the polypeptide has the following nucleic acid sequence:

tccaccaaggcccatecggtttcccgctagcgccctgtccaggagcacctccgagagc
acagccgcctgggtcgaa
tcaaggacttccccgaaccgggtgacggtgtcgtaactcaggcgccctgaccagcg
gcgtgcacaccttccgggtgtc
ctacagtctcaggactctactccctcagcagcgtggaccgtgcctccagcagcttg
ggcacaagaacctacacctgcaac
gtatgtcacaaggcccgaaacaccaagggtggacaagagagttgagtccaaatatggtccc
ccatggccaccctggcccgacca
cctgagtttctggggggaccatcgtcttcgttcccccggaaaacccaaggacactctc
atgtatccctggggaccctgtgggtc
gtggcgtgggtggacgtggccaggaaaggccccgggtccagttcaactggtaactggta
tggcgtggagggtgcataatgcga
agacaaggccggggaggaggcagttcaacacagcacgtaccgtggtcagcgtccatcacc
tcctgcacaggactggctgaa
cggcaaggagttacaagggtctccaacaaaggcctccgtccatcgagaaaac
catctccaaagccaaaggcca

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gccccggagggccacagggttacaccctgccccatcccaggaggagatgaccaagaacca
ggtcagcctgacctgcctggc
aaaggcttctaccccaagcgacatcgccgtggagtggagagacaatggcagccggagaac
aactacaagaccacgcctccc
gtgtggactccgcacggctctttctctacagcaggctaaccgtggacaagagcagg
ttgcaggaggggaatgttcttcatgc
tccgtgtatgcatgaggctctgcacaaccactacacacagaagagcctccctgtctctg
ggtaaatga.

40. A polynucleotide encoding a heterologous fusion protein of any one of Claims 1 through 389.

41. A vector comprising the polynucleotide of Claim 40.

42. A host cell comprising the vector of Claim 41.

43. A host cell expressing at least one heterologous fusion protein of any one of Claims 1 through 39.

44. The host cell of Claim 43 wherein said host cell is a CHO cell.

45. A process for producing a heterologous fusion protein comprising the steps of transcribing and translating a polynucleotide of Claim 40 under conditions wherein the heterologous fusion protein is expressed in detectable amounts.

46. A method for increasing neutrophil levels in a mammal comprising the administration of a therapeutically effective amount of the heterologous fusion protein of any one of Claims 27 through 36.

47. The use of the heterologous fusion protein as claimed in any one of Claims 22 through 29 for the manufacture of a medicament for the treatment of patients with insufficient circulating neutrophil levels.

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48. Use of a heterologous fusion protein of any one of Claims 22 through 29 as a medicament.

49. Use of a heterologous fusion protein of any one of Claims 22 through 29 for the treatment of patients with insufficient circulating neutrophil levels.

50. A pharmaceutical formulation adapted for the treatment of patients with insufficient neutrophil levels comprising a heterologous fusion protein of any one of Claims 22 through 29.

51. A heterologous fusion protein as hereinbefore described with reference to any one of the Examples.

SEQUENCE LISTING

<110> Beals, John

Kuchibhotla, Uma

<120> HETEROLOGOUS G-CSF FUSION PROTEINS

<130> P-15648

<160> 66

<170> PatentIn version 3.1

<210> 1

<211> 174

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<223> Xaa at position 58 is Trp or Asn;

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<223> Xaa at position 60 is Pro, Thr, Asn, or Ser;

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<222> (61)..(61)

<223> Xaa at position 61 is Leu, or any other amino acid except Pro;

<220>

<221> MISC_FEATURE

<222> (62)..(62)

<223> Xaa at position 62 is Ser or Thr;

<220>

<221> MISC_FEATURE

<222> (63)..(63)

<223> Xaa at position 63 Ser or Asn;

<220>

<221> MISC_FEATURE

<222> (64)..(64)

<223> Xaa at position 64 is Cys or any other amino acid except Pro;

<220>

<221> MISC_FEATURE

<222> (65)..(65)

<223> Xaa at position 65 is Pro, Ser, or Thr;

<220>

<221> MISC_FEATURE

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<222> (66)..(66)

<223> Xaa at position 66 is Ser or Thr;

<220>

<221> MISC_FEATURE

<222> (67)..(67)

<223> Xaa at position 67 is Gln or Asn;

<220>

<221> MISC_FEATURE

<222> (68)..(68)

<223> Xaa at position 68 is Ala or any other amino acid except Pro;

<220>

<221> MISC_FEATURE

<222> (69)..(69)

<223> Xaa at position 69 is Leu, Thr, or Ser;

<220>

<221> MISC_FEATURE

<222> (93)..(93)

<223> Xaa at position 93 is Glu or Asn;

<220>

<221> MISC_FEATURE

<222> (94)..(94)

<223> Xaa at position 94 is Gly or any other amino acid except Pro;

<220>

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<221> MISC_FEATURE
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<223> Xaa at position 95 is Ile, Asn, Ser, or Thr;

<220>
<221> MISC_FEATURE
<222> (97)..(97)
<223> Xaa at position 97 is Pro, Ser, Thr, or Asn;

<220>
<221> MISC_FEATURE
<222> (133)..(133)
<223> Xaa at position 133 is Thr or Asn;

<220>
<221> MISC_FEATURE
<222> (134)..(134)
<223> Xaa at position 134 is Gln or any other amino acid except Pro;

<220>
<221> MISC_FEATURE
<222> (135)..(135)
<223> Xaa at position 135 is Gly, Ser, or Thr;

<220>
<221> MISC_FEATURE
<222> (141)..(141)
<223> Xaa at position 141 is Ala or Asn;

<220>

<221> MISC_FEATURE

<222> (142)..(142)

<223> Xaa at position 142 is Ser or any other amino acid except Pro;

<220>

<221> MISC_FEATURE

<222> (143)..(143)

<223> Xaa at position 143 is Ala, Ser, or Thr.

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Xaa	Leu	Glu	Gln	Val	Arg	Lys	Ile	Gln	Gly	Asp	Gly	Ala	Ala	Leu	Gln
							20		25					30	

Glu	Lys	Leu	Cys	Xaa	Xaa	Xaa	Lys	Leu	Cys	His	Pro	Glu	Glu	Leu	Val
							35		40				45		

Leu	Leu	Gly	His	Ser	Leu	Gly	Ile	Xaa							
							50		55				60		

Xaa	Xaa	Xaa	Xaa	Xaa	Gln	Leu	Ala	Gly	Cys	Leu	Ser	Gln	Leu	His	Ser
							65		70			75		80	

Gly	Leu	Phe	Leu	Tyr	Gln	Gly	Leu	Leu	Gln	Ala	Leu	Xaa	Xaa	Xaa	Ser
							85		90			95			

Xaa	Glu	Leu	Gly	Pro	Thr	Leu	Asp	Thr	Leu	Gln	Leu	Asp	Val	Ala	Asp
							100		105			110			

Phe	Ala	Thr	Thr	Ile	Trp	Gln	Gln	Met	Glu	Glu	Leu	Gly	Met	Ala	Pro
							115		120			125			

Ala	Leu	Gln	Pro	Xaa	Xaa	Xaa	Ala	Met	Pro	Ala	Phe	Xaa	Xaa	Xaa	Phe
							130		135			140			

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Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe
145 150 155 160

Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro
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<213> Artificial Sequence

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acactgcgcg tggacgttccg cgcacgggc acacccgggtt ggaacctgtt tgacgttccg 660
ctgcacggcgtt ttgttgcac caccatctgg cggcaggatgg aagaactggg aatggccct 720
aaacgggtgtt ggttagaccgtt cgttcttacccctt ctgttccctt accggggggac cctgtcgaccc 780
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<213> Artificial Sequence

<220>

<223> synthetic construct

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<210> 4

<211> 1044

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<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

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<212> DNA

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<220>

<223> synthetic construct

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cgtcgcgagg tcgagaagct gtgtgccacc tacaagctgt gccaccccgaa ggagctggtg 240

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211 1044

5312 DNA

<213> Artificial Sequence

53303

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54003-8

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<211> 1044

<212> DNA

<213> Artificial Sequence

<220>

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cagaattccg tggAACGGGT CGGG 1044

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<212> DNA
<213> Artificial Sequence

<220>

<223> synthetic construct

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<210> 14

<211> 1044

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

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<210> 15

<211> 1044

<212> DNA

<213> Artificial Sequence

<220>

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<223> synthetic construct

<400> 15
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<211> 1044

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

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<211> 1762

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

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aatttgcata	aatcacttca	taccctttt	ggagacaaat	tatgcacagt	tgcactt	tt	240
cgtgaaacct	atggtgaaat	ggctgactgc	tgtgcacaaac	aagaacctga	gagaatgaa	aa	300

tgcttcttc aacacaaga tgacaaccca aacctccccc gattggtag accagaggtt 360
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gaaattgcca gaagacatcc ttactttat gccccggAAC tcctttctt tgctaaaagg 480
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<210> 18

<211> 232

<212> PRT

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 18

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Ala	Pro	Glu	Lys	Gly	Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	Lys	Pro
						20			25				30		

Lys	Asp	Thr	Lys	Met	Ile	Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val
				35				40				45			

Val	Asp	Val	Ser	His	Glu	Asp	Pro	Glu	Val	Lys	Phe	Asn	Trp	Tyr	Val
				50			55				60				

Asp	Gly	Val	Glu	Val	His	Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	Glu	Gln
					65			70			75			80	

Tyr	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser	Val	Leu	Thr	Val	Leu	His	Gln
					85				90				95		

Asp	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Ala
				100				105				110			

Leu	Pro	Ala	Pro	Ile	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	Gln	Pro
				115				120				125			

Arg	Glu	Pro	Gln	Val	Tyr	Thr	Leu	Pro	Pro	Ser	Arg	Glu	Glu	Met	Thr
				130				135			140				

Lys	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	Pro	Ser
				145				150			155			160	

Asp	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn	Gly	Gln	Pro	Glu	Asn	Asn	Tyr
				165				170			175				

Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr
					180				185			190			

Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe
195 200 205

Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys
210 215 220

Ser Leu Ser Leu Ser Pro Gly Lys
225 230

<210> 19

<211> 229

<212> PRT

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 19

Glu Ser Lys Tyr Gly Pro Pro Cys Pro Pro Cys Pro Ala Pro Glu Phe
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Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr
20 25 30

Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val
35 40 45

Ser Gln Glu Asp Pro Glu Val Gln Phe Asn Trp Tyr Val Asp Gly Val
50 55 60

Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Phe Asn Ser
65 70 75 80

Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu
85 90 95

Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Gly Leu Pro Ser
100 105 110

Ser Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro
115 120 125

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Gln Val Tyr Thr Leu Pro Pro Ser Gln Glu Glu Met Thr Lys Asn Gln
130 135 140

Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala
145 150 155 160

Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr
165 170 175

Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Arg Leu
180 185 190

Thr Val Asp Lys Ser Arg Trp Gln Glu Gly Asn Val Phe Ser Cys Ser
195 200 205

Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser
210 215 220

Leu Ser Leu Gly Lys
225

<210> 20

<211> 585

<212> PRT

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 20

Asp Ala His Lys Ser Glu Val Ala His Arg Phe Lys Asp Leu Gly Glu
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Glu Asn Phe Lys Ala Leu Val Leu Ile Ala Phe Ala Gln Tyr Leu Gln
20 25 30

Gln Cys Pro Phe Glu Asp His Val Lys Leu Val Asn Glu Val Thr Glu
35 40 45

Phe Ala Lys Thr Cys Val Ala Asp Glu Ser Ala Glu Asn Cys Asp Lys
50 55 60

Ser Leu His Thr Leu Phe Gly Asp Lys Leu Cys Thr Val Ala Thr Leu
65 70 75 80

Arg Glu Thr Tyr Gly Glu Met Ala Asp Cys Cys Ala Lys Gln Glu Pro
85 90 95

Glu Arg Asn Glu Cys Phe Leu Gln His Lys Asp Asp Asn Pro Asn Leu
100 105 110

Pro Arg Leu Val Arg Pro Glu Val Asp Val Met Cys Thr Ala Phe His
115 120 125

Asp Asn Glu Glu Thr Phe Leu Lys Lys Tyr Leu Tyr Glu Ile Ala Arg
130 135 140

Arg His Pro Tyr Phe Tyr Ala Pro Glu Leu Leu Phe Phe Ala Lys Arg
145 150 155 160

Tyr Lys Ala Ala Phe Thr Glu Cys Cys Gln Ala Ala Asp Lys Ala Ala
165 170 175

Cys Leu Leu Pro Lys Leu Asp Glu Leu Arg Asp Glu Gly Lys Ala Ser
180 185 190

Ser Ala Lys Gln Arg Leu Lys Cys Ala Ser Leu Gln Lys Phe Gly Glu
195 200 205

Arg Ala Phe Lys Ala Trp Ala Val Ala Arg Leu Ser Gln Arg Phe Pro
210 215 220

Lys Ala Glu Phe Ala Glu Val Ser Lys Leu Val Thr Asp Leu Thr Lys
225 230 235 240

Val His Thr Glu Cys Cys His Gly Asp Leu Leu Glu Cys Ala Asp Asp
245 250 255

Arg Ala Asp Leu Ala Lys Tyr Ile Cys Glu Asn Gln Asp Ser Ile Ser
260 265 270

Ser Lys Leu Lys Glu Cys Cys Glu Lys Pro Leu Leu Glu Lys Ser His
275 280 285

Cys Ile Ala Glu Val Glu Asn Asp Glu Met Pro Ala Asp Leu Pro Ser
290 295 300

Leu Ala Ala Asp Phe Val Glu Ser Lys Asp Val Cys Lys Asn Tyr Ala
305 310 315 320

Glu Ala Lys Asp Val Phe Leu Gly Met Phe Leu Tyr Glu Tyr Ala Arg
325 330 335 320

Arg His Pro Asp Tyr Ser Val Val Leu Leu Leu Arg Leu Ala Lys Thr
340 345 350

Tyr Glu Thr Thr Leu Glu Lys Cys Cys Ala Ala Ala Asp Pro His Glu
355 360 365

Cys Tyr Ala Lys Val Phe Asp Glu Phe Lys Pro Leu Val Glu Glu Pro
370 375 380

Gln Asn Leu Ile Lys Gln Asn Cys Glu Leu Phe Glu Asn Leu Gly Glu
385 390 395 400

Tyr Lys Phe Gln Asn Ala Leu Leu Val Arg Tyr Thr Lys Lys Val Pro
405 410 415

Gln Val Ser Thr Pro Thr Leu Val Glu Val Ser Arg Asn Leu Gly Lys
420 425 430

Val Gly Ser Lys Cys Cys Lys His Pro Glu Ala Lys Arg Met Pro Cys
435 440 445

Ala Glu Asp Tyr Leu Ser Val Val Leu Asn Gln Leu Cys Val Leu His
450 455 460

Glu Lys Thr Pro Val Ser Asp Arg Val Thr Lys Cys Cys Thr Glu Ser
465 470 475 480

Leu Val Asn Arg Arg Pro Cys Phe Ser Ala Leu Glu Val Asp Glu Thr
485 490 495

Tyr Val Pro Lys Glu Phe Asn Ala Glu Thr Phe Thr Phe His Ala Asp
500 505 510

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Ile Cys Thr Leu Ser Glu Lys Glu Arg Gln Ile Lys Lys Gln Thr Ala
515 520 525

Leu Val Glu Leu Val Lys His Lys Pro Lys Ala Thr Lys Glu Gln Leu
530 535 540

Lys Ala Val Met Asp Asp Phe Ala Ala Phe Val Glu Lys Cys Cys Lys
545 550 555 560

Ala Asp Asp Lys Glu Thr Cys Phe Ala Glu Glu Gly Lys Lys Leu Val
565 570 575

Ala Ala Ser Gln Ala Ala Leu Gly Leu
580 585

<210> 21

<211> 703

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

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ccccgtgtgg actccgacgg ctcccttcttc ctctatagca agtcaccgtt ggacaaggacg 600
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<210> 22
<211> 981
<212> DNA
<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 22	
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ctctactccc tcagcagcgt ggtgaccgtg ccctcccgca gcttgggcac gaagacactac	240
acctgcaacg tagatcacaa gcccagcaac accaagggtgg acaagagagt tgagtccaaa	300
tatggtcccc catgccccacc ctgccccagca cctgagttcc tggggggacc atcagtctt	360
ctgttccccca caaaacccaa ggacactctc atgatctccc ggacccctga ggtcacgtgc	420
gtgggtgtgg acgtgagcca ggaagacccc gaggtccagt tcaactggta cgtggatggc	480
gtggagggtgc ataatgccaa gacaaggccg cgggaggagc agttcaacacg caccgtaccgt	540
gtggtcagecg tcctcaccgt cctgcaccag gactggctga acggcaaggg gtacaagtgc	600
aagggtctcca acaaaggccct cccgtctccc atcgagaaaa ccattctccaa agccaaagg	660
cagccccgag agccacaggt gtacaccctg ccccccattcc aggaggagat gaccaagaac	720
cagggtcagcc tgacctgcct ggtcaaaaggc ttcttacccca gcgacatcgcc cgtgagtg	780
gagagcaatg ggcagccgga gaacaactac aagaccacacgc ctcccggtgt ggactccgac	840
ggctctttct tcctctacag caggctaaacc gtggacaaga gcagggtggca ggagggaaat	900
gtcttctcat gtcctgtgtat gcatgaggct ctgcacaacc actacacaca gaagagccctc	960
tccctgtctc tgggtaaatg a	981

<210> 23

<211> 406

<212> PRT

213 Artificial Sequence

220

<223> synthetic construct

<400> 23

Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Leu Lys
1 5 10 15

Ala Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln
20 25 30

Glu Lys Leu Cys Ala Thr Tyr Lys Leu Cys His Pro Glu Glu Leu Val
35 40 45

Leu Leu Gly His Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser Cys
50 55 60

Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser
65 70 75 80

Gly Leu Phe Leu Tyr Gln Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser
85 86 87 88 89 90 91 92 93 94 95

Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp
 100 105 110

Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro
 115 120 125

Ala Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe
120 125 130

Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe
145 150 155 160

Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro Glu Pro
155 156 157 158 159

Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu

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Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp
195 200 205

Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp
210 215 220

Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly
225 230 235 240

Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn
245 250 255

Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp
260 265 270

Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro
275 280 285

Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu
290 295 300

Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn
305 310 315 320

Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile
325 330 335

Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr
340 345 350

Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys
355 360 365

Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys
370 375 380

Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu
385 390 395 400

Ser Leu Ser Pro Gly Lys
405

<211> 403

52128 PRT

<213> Artificial Sequence

1330

<223> synthetic construct

-400- 34

Ala Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln
20 25 30

Glu Lys Leu Cys Ala Thr Tyr Lys Leu Cys His Pro Glu Glu Leu Val
35 40 45

Leu Leu Gly His Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser Cys
50 55 60

Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser
65 70 75 80

Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser
 ²⁵ ³⁰ ³⁵

Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp
189 190 191 192 193 194 195 196 197 198 199 200 201 202 203

Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro
115 120 125

Ala Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe
120 125 130

Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe
155 156 157 158 159 160

Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro Glu Ser
165 170 175

Lys Tyr Gly Pro Pro Cys Pro Ser Cys Pro Ala Pro Glu Phe Leu Gly
180 185 190

Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met
195 200 205

Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser Gln
210 215 220

Glu Asp Pro Glu Val Gln Phe Asn Trp Tyr Val Asp Gly Val Glu Val
225 230 235 240

His Asn Ala Lys Thr Lys Pro Arg Glu Gln Phe Asn Ser Thr Tyr
245 250 255

Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly
260 265 270

Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Gly Leu Pro Ser Ser Ile
275 280 285

Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val
290 295 300

Tyr Thr Leu Pro Pro Ser Gln Glu Glu Met Thr Lys Asn Gln Val Ser
305 310 315 320

Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu
325 330 335

Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro
340 345 350

Val Leu Asp Ser Asp Gly Ser Phe Phe Tyr Ser Arg Leu Thr Val
355 360 365

Asp Lys Ser Arg Trp Gln Glu Gly Asn Val Phe Ser Cys Ser Val Met
370 375 380

His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser
385 390 395 400

Leu Gly Lys

<210> 25

<211> 500

<212> PRT

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 25

Thr	Pro	Leu	Gly	Pro	Ala	Ser	Ser	Leu	Pro	Gln	Ser	Phe	Leu	Leu	Lys
1					5				10					15	

Ala	Leu	Glu	Gln	Val	Arg	Lys	Ile	Gln	Gly	Asp	Gly	Ala	Ala	Leu	Gln
				20				25				30			

Glu	Lys	Leu	Cys	Ala	Thr	Tyr	Lys	Leu	Cys	His	Pro	Glu	Glu	Leu	Val
35						40					45				

Leu	Leu	Gly	His	Ser	Leu	Gly	Ile	Pro	Trp	Ala	Pro	Leu	Ser	Ser	Cys
50					55				60						

Pro	Ser	Gln	Ala	Leu	Gln	Leu	Ala	Gly	Cys	Leu	Ser	Gln	Leu	His	Ser
65					70				75			80			

Gly	Leu	Phe	Leu	Tyr	Gln	Gly	Leu	Leu	Gln	Ala	Leu	Glu	Gly	Ile	Ser
			85					90				95			

Pro	Glu	Leu	Gly	Pro	Thr	Leu	Asp	Thr	Leu	Gln	Leu	Asp	Val	Ala	Asp
	100						105					110			

Phe	Ala	Thr	Thr	Ile	Trp	Gln	Gln	Met	Glu	Glu	Leu	Gly	Met	Ala	Pro
	115					120					125				

Ala	Leu	Gln	Pro	Thr	Gln	Gly	Ala	Met	Pro	Ala	Phe	Ala	Ser	Ala	Phe
	130				135				140						

Gln	Arg	Arg	Ala	Gly	Gly	Val	Leu	Val	Ala	Ser	His	Leu	Gln	Ser	Phe
145					150				155			160			

Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro Gly Gly
165 170 175

Gly Gly Ser Gly Gly Gly Ser Gly Gly Gly Ser Asp Ala His
180 185 190

Lys Ser Glu Val Ala His Arg Phe Lys Asp Leu Gly Glu Glu Asn Phe
195 200 205

Lys Ala Leu Val Leu Ile Ala Phe Ala Gln Tyr Leu Gln Gln Cys Pro
210 215 220

Phe Glu Asp His Val Lys Leu Val Asn Glu Val Thr Glu Phe Ala Lys
225 230 235 240

Thr Cys Val Ala Asp Glu Ser Ala Glu Asn Cys Asp Lys Ser Leu His
245 250 255

Thr Leu Phe Gly Asp Lys Leu Cys Thr Val Ala Thr Leu Arg Glu Thr
260 265 270

Tyr Gly Glu Met Ala Asp Cys Cys Ala Lys Gln Glu Pro Glu Arg Asn
275 280 285

Glu Cys Phe Leu Gln His Lys Asp Asp Asn Pro Asn Leu Pro Arg Leu
290 295 300

Val Arg Pro Glu Val Asp Val Met Cys Thr Ala Phe His Asp Asn Glu
305 310 315 320

Glu Thr Phe Leu Lys Lys Tyr Leu Tyr Glu Ile Ala Arg Arg His Pro
325 330 335

Tyr Phe Tyr Ala Pro Glu Leu Leu Phe Phe Ala Lys Arg Tyr Lys Ala
340 345 350

Ala Phe Thr Glu Cys Cys Gln Ala Ala Asp Lys Ala Ala Cys Leu Leu
355 360 365

Pro Lys Leu Asp Glu Leu Arg Asp Glu Gly Lys Ala Ser Ser Ala Lys
370 375 380

Gln Arg Leu Lys Cys Ala Ser Leu Gln Lys Phe Gly Glu Arg Ala Phe
385 390 395 400

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Lys Ala Trp Ala Val Ala Arg Leu Ser Gln Arg Phe Pro Lys Ala Glu
405 410 415

Phe Ala Glu Val Ser Lys Leu Val Thr Asp Leu Thr Lys Val His Thr
420 425 430

Glu Cys Cys His Gly Asp Leu Leu Glu Cys Ala Asp Asp Arg Ala Asp
435 440 445

Leu Ala Lys Tyr Ile Cys Glu Asn Gln Asp Ser Ile Ser Ser Lys Leu
450 455 460

Lys Glu Cys Cys Glu Lys Pro Leu Leu Glu Lys Ser His Cys Ile Ala
465 470 475 480

Glu Val Glu Asn Asp Glu Met Pro Ala Asp Leu Pro Ser Leu Ala Ala
485 490 495

Asp Phe Val Glu
500

<210> 26

<211> 69

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 26
gtaaagcttgc gtcgacacgcta gggcgccgc gccatggccg gacctggccac ccagagcccc
atgaagctg 60
69

<210> 27

<211> 61

<212> DNA

<213> Artificial Sequence

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PCT/US03/03120

36/48

<220>

<223> synthetic construct

<400> 27
ggggcaggga gctggctggg cccagtggag tggcttcctg cactgtccag agtgcactgt 60
g 61

<210> 28

<211> 59

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 28
ggacagtgtca ggaagccact ccactgggcc cagccagctc cctgccccag agcttcctg 59

<210> 29

<211> 72

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 29
gaacctcgag gatcctcatt agggctgggc aagggtgcctt aagacgcgggt acgacacctc 60
caggaagctc tg 72

<210> 30

<211> 69

<212> DNA

<213> Artificial Sequence

WO 03/076567

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37/48

<220>

<223> synthetic construct

<400> 30
gttaagcttgc gtcgacgcta gggcgccccc gccatggccg gacctgccac ccagagcccc 60
atgaagctg 69

<210> 31

<211> 57

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 31
gctctaaaggc cttgagcagg aagctctggg gcagggagct cgctggggccc agtggag 57

<210> 32

<211> 53

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 32
ggcccccagcg agctccctgc cccagagctt cctgctcaag gccttagagc aag 53

<210> 33

<211> 72

<212> DNA

<213> Artificial Sequence

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38/48

<220>

<223> synthetic construct

<400> 33	gacacatcgag gatcctcatt agggctgggc aagggtgcctt aagacgcggc acgacaccc	60
caggaagctc tg		72

<210> 34

<211> 69

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 34	gttaaagcttc gtcgacgcta gggcgccccc gccatggccg gacctgccac ccagagcccc	60
atgaagctg		69

<210> 35

<211> 61

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 35	gtccggcggc cactagttcc tcgggggtggc acagcttggt gggttacac agcttctct	60
g		61

<210> 36

<211> 66

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 36
ggcgccgcg tccaggagaa gctgtgttaac accaccaagc tgtgccaccc cgaggaacta 60
gtgctg

<210> 37

<211> 72

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 37
gaacctcgag gatcctcatt agggctggc aagggtgcctt aagacgcggc acgacacacctc 60
caggaagctc tg 72

<210> 38

<211> 69

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 38
gtaagcttgc gtgcacgcta gggcgccgc gccatggccg gacctgccac ccagagcccc 60
atgaagctg 69

<210> 39

<211> 61

<212> DNA

WO 03/076567

PCT/US03/03120

40/48

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 39
ggccggcgct ggaaaggcgt ggcgaaggcc ggcattggcg tctggttggg ctgcaggca 60
g 61

<210> 40

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 40
ggccccctgcc ctgcagccca accagaccgc catgcccggcc ttccggcagcg ctttccagcg 60

<210> 41

<211> 72

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 41
gaacacctcgag gatcctcatt agggctgggc aaggtgcctt aagacgcggc acgacacactc 60
caggaagactc tg 72

<210> 42

<211> 69

<212> DNA

WO 03/076567

PCT/US03/03120

41/48

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 42
gtaaagttgc gtcgacgcta gggcgccgc gccatggccg gacctggcac ccagagcccc 60
atgaagctg 69

<210> 43

<211> 68

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 43
gccccggcgt ggaaggtaga gttgaaggcc ggcatggcac cctgggtggg ctgaagagca 60
ggggccat 68

<210> 44

<211> 74

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 44
ggaaatggcc cctgcttcc agccccaccca gggtgccatg cgggccttca actctacatt 60
ccagcgccgg gcag 74

<210> 45

<211> 72

WO 03/076567

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42/48

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 45
gaacctcgag gatcctcatt agggctgggc aagggtgcctt aagacgcggt acgacacacctc 60
caggaagtc tg 72

<210> 46

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 46
gcttagccgcg cgccaccatg 20

<210> 47

<211> 33

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 47
gctcagggtta cggttaacgta tgccccagaga gtg 33

<210> 48

<211> 30

<212> DNA

WO 03/076567

PCT/US03/03120

43/48

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 48

gggcgtcggtt aacgcttaccc tgagcagctg

30

<210> 49

<211> 27

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 49

gactcgagga tcctcattag ggctggg

27

<210> 50

<211> 38

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 50

gctagccggcg cgccaccatg gccggacctg ccacccag

38

<210> 51

<211> 45

<212> DNA

<213> Artificial Sequence

WO 03/076567

PCT/US03/03120

44/48

<220>

<223> synthetic construct

<400> 51
caagcagccg gccagctggg tggcggttgc ggggcagctg cttag 45

<210> 52

<211> 37

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 52
gccccagcaa cgccacccag ctggccggct gctttag 37

<210> 53

<211> 47

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 53
gactcgagga tcctcattag ggctggccaa ggtgccttaa gacgcgg 47

<210> 54

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

WO 03/076567

PCT/US03/03120

45/48

<400> 54
gctagcgccg cgcaccatg

20

<210> 55

<211> 27

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 55
ggggcaacta gtcaggtag cccaggg

27

<210> 56

<211> 27

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 56
gctaacctga ctatgtccc cagccag

27

<210> 57

<211> 27

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 57
gactcgagga tcctcattag ggctggg

27

WO 03/076567

PCT/US03/03120

46/48

<210> 58
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> synthetic construct
<400> 58
gttagcggcg cgccaccatg

20

<210> 59
<211> 25
<212> DNA
<213> Artificial Sequence

<220>
<223> synthetic construct
<400> 59
ggtgcaattg ctcagggag cccag

25

<210> 60
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> synthetic construct
<400> 60
gcaattgcac cagccaggcc ctg

23

<210> 61
<211> 27

WO 03/076567

PCT/US03/03120

47/48

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 61

gactcgagga tcctcattag ggctggg

27

<210> 62

<211> 38

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 62

gctagcggcg cgccaccatg gccggacctg ccacccag

38

<210> 63

<211> 37

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 63

ccggacttgtt cccgttcagg gcctgcagga gcccctg

37

<210> 64

<211> 35

<212> DNA

<213> Artificial Sequence

WO 03/076567

PCT/US03/03120

48/48

<220>

<223> synthetic construct

<400> 64

gaacgggacc agtccggagt tgggtccccac cttagg

35

<210> 65

<211> 47

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 65

gactcgagga tcctcattag ggctggcaa ggtgccttaa gacgcgg

47

<210> 66

<211> 36

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 66

gtcgacgcta gcggcgcc accatggccg gacctg

36